



The Protist Ribosomal Reference databases

Daniel Vaultot

BIO9905MERG1 course - 2025-04-10



CNRS • SORBONNE UNIVERSITÉ
Station Biologique
de Roscoff



UiO : **University of Oslo**

Roscoff



La Station Biologique de Roscoff



- 1872 - Antoine Lacaze-Duthiers (150 years ago)
- CNRS and Sorbonne Université
- Staff: 350
- Students : 1,000-2,000 per year

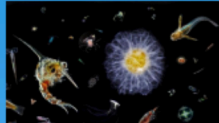


Ecologie of Marine Plankton team

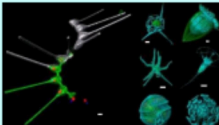
- Scientists: 14
- Staff: ~ 40
- Research themes:
 - Viruses
 - Bacteria
 - Cyanobacteria
 - Symbioses
 - Parasitism

ECOLOGY OF MARINE PLANKTON (ECOMAP)

PERMANENT STAFF

Anne-Claire BAUDOUX CRCN CNRS Marine viruses: ecology, diversity and evolution	Aurélien CHAMBOUVET CRCN CNRS Diversity and Ecology of metazoan parasitic protists	Laurence GARCZAREK DR1 CNRS Genetic and functional diversity of marine cyanobacteria	Benjamin BAILLEUL CRCN CNRS Photophysiology of phytoplankton 49% ECOMAP 51% IBPC	Juliana BERNARDES MC SU Machine learning in marine ecology
Colomban DE VARGAS DR1 CNRS & FR GOSSE Plankton system biology	Laure GUILLOU DR1 CNRS DU UMR symbiosis and parasitism	Christian JEANTHON DR2 CNRS Ecology of marine bacteria and their interactions with phytoplankton	Fabrice NOT DR2 CNRS Plankton symbiosis: diversity, ecology & evolution	Frédéric PARTENSKY DR1 CNRS Function and evolution of cyanobacterial light-harvesting complexes
Nathalie SIMON MC SU Phytoplankton biodiversity and temporal dynamics	Christophe SIX MC SU Adaptation of phytoplankton to temperature	Daniel VAULOT DRCE CNRS Emeritus Diversity and ecology of eukaryotic picoplankton	Noriko YAMADA CRCN CNRS Evolutionary developmental processes of plastids in "dinotom" dinoflagellates	
Estelle BIGEARD IE CNRS Marine eukaryotic parasites & viruses Lab Management (LabCollector)	Florence LE GALL IEHC CNRS Marine plankton Lab management Permanent Training	Camille POIRIER IE CNRS Flow cytometry & Lab Management 90% ECOMAP 10% RECYF	Charlotte BERTHELIER IE CNRS Bioanalyses of marine plankton	Nicolas HENRY IR FR2424 Plankton eco- morphogenesis
Morgane RATIN IR CNRS Mol. biology & genetics of cyanobacteria Security & safety	Sarah ROMAC IE CNRS Molecular ecology of marine protists Lab management		Fabienne RIGAUD IR FR2424 PI	

POST-DOCS, PHD STUDENTS & TEMPORARY STAFF

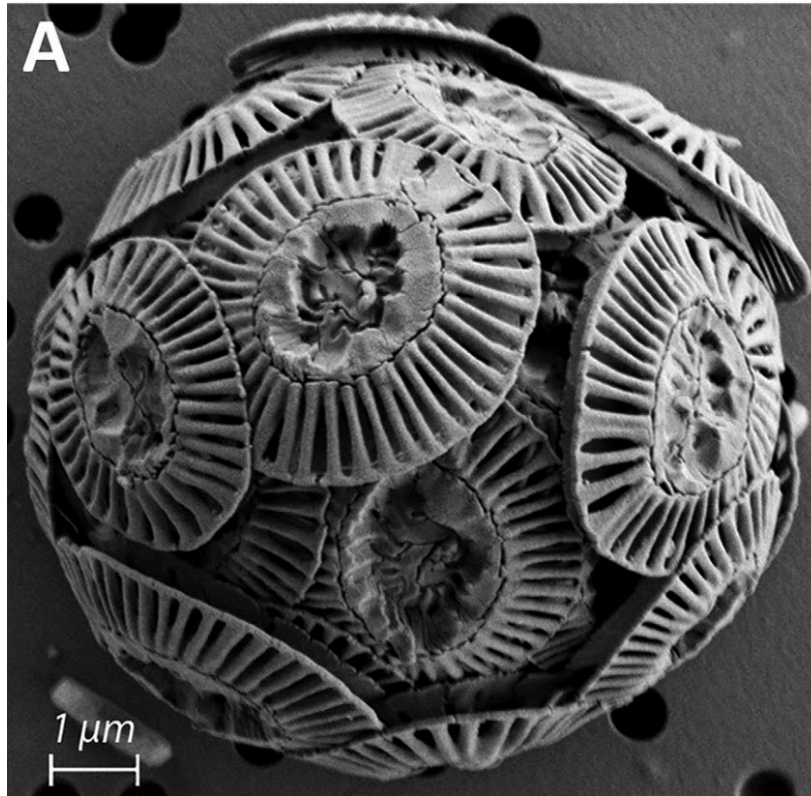
Emile FAURE Post-Doc Metagenomics of marine picocyanobacteria	Valeria JIMENEZ Post-Doc Dinoflagellate- diatom symbiosis	Natalia LLOPIS MONFERRER Post-Doc Silicification in Rhizaria	Ian PROBERT IRHC SU FR2424 RCC Manager	Priscillia GOURVIL IR CNRS FR2424 Curator of the RCC
Francesco MATTEI Post-Doc Interactions between diatoms and bacteria	Morgan SMITS Post-Doc Chemical interactions between host and parasite	Elisa CHAILLER PhD student Role of pathobiome on parasitic infections	Charles BACHY IR CNRS FR2424 RCC R&D manager	Martin GACHENOT AI SU FR2424 RCC Strain maintenance & Flow cyto. platform (RECYF)
Emmanuelle JAOUEN PhD student Marine viruses	Perrine KERGOAT PhD student Niche adaptation ecology of marine cyanobacteria	Heliaz LE BAYON PhD student Molecular ecology of shellfish parasites	Sarah GARRIC IE SU FR2424 Characterization of cryptophytes	Michele GREGO AI SU FR2424 RCC Cryo- preservation
Morgane GUILLAM IE SU Bougainville project Biological engineer	Erwan LEGEAY IE SU Environmental Genomics	Julie LEPETIT AI Lipidomics of dinoflagellates	Katell HERVEOU AI SU FR2424 Strain maintenance & ATLASEA	Julie JOANNIC AI SU FR2424 Molecular biology of phytoplankton
Pauline NOGARET AI Marine viruses			Marie WALDE IR SU FR2424 Advanced imaging of marine plankton	

ROSCOFF CULTURE COLLECTION



Roscoff Culture Collection

- Largest marine algal collection
- 9,500 strains distributed



<https://roscoff-culture-collection.org/>



HOME STRAINS KNOWLEDGEBASE ORDERING / DEPOSITING ABOUT RCC CONTACT MY CART

Login

Register

Roscoff Culture Collection

1 Frequently Asked Questions

Welcome to the Roscoff Culture Collection

Since JULY 2021, we ONLY accept payments by credit card via a secured banking system : PayBox.

The Roscoff Culture Collection (RCC) maintains and distributes approximately 5000 strains of marine microalgae, macroalgae, protists, bacteria and viruses. It is located at the [Station Biologique in Roscoff \(France\)](#) which is run by [Sorbonne University](#) and the [CNRS](#). The RCC is part of the European infrastructure [EMBRIC](#) and the French national infrastructure [EMBRIC-France](#).

In addition to services offered for public and private laboratories, the RCC supports schools in carrying out practical work. Pour plus d'information sur la fourniture de souches pour les colleges et lycées [cliquez ici](#).

Quick Search

RCC Number

Class

Genus

- Any -

- Any -

Search

Clear

Strain catalogue

Strain catalogue

Strain Map

Media recipes

Services offered

How to order

Cyanobacteria

Green algae

Haptophytes

Diatoms

Dinoflagellates

RCC1020

RCC4076

RCC5268

RCC5605

RCC5711

Strains maintained

5,894

Strains distributed

1,878

Strains cryopreserved

973

Species available

Strain pictures

2,882

Sequences available

6,514

Papers available

447

Tweets by @RCC_algae

Roscoff Culture Collection

@RCC_algae

New using an RCC strain RCC1359

High-value lipids accumulation by Pavlova pinguis as a response to nitrogen-induced changes by Tomásia Fernandes and Nereida Cordeiro. [sciencedirect.com/science/articl...](#)

High-value lipids a...

The challenges of e...

sciencedirect.com

Roscoff Culture Collection

@RCC_algae

New paper on Chlorococcophyceae, dominant green algae in the moderately oligotrophic ocean using 3 RCC strains. Ebenezer et al. Elemental and macromolecular composition of the marine Chlorococcophyceae, a major group of oceanic photosynthetic picoeukaryotes [doi.org/10.1002/no.12...](#)

Roscoff Culture Collection

Retweeted

BCCM Collections

@BCCMCollections

Sharing is caring, and we care about your biological materials! 🧪

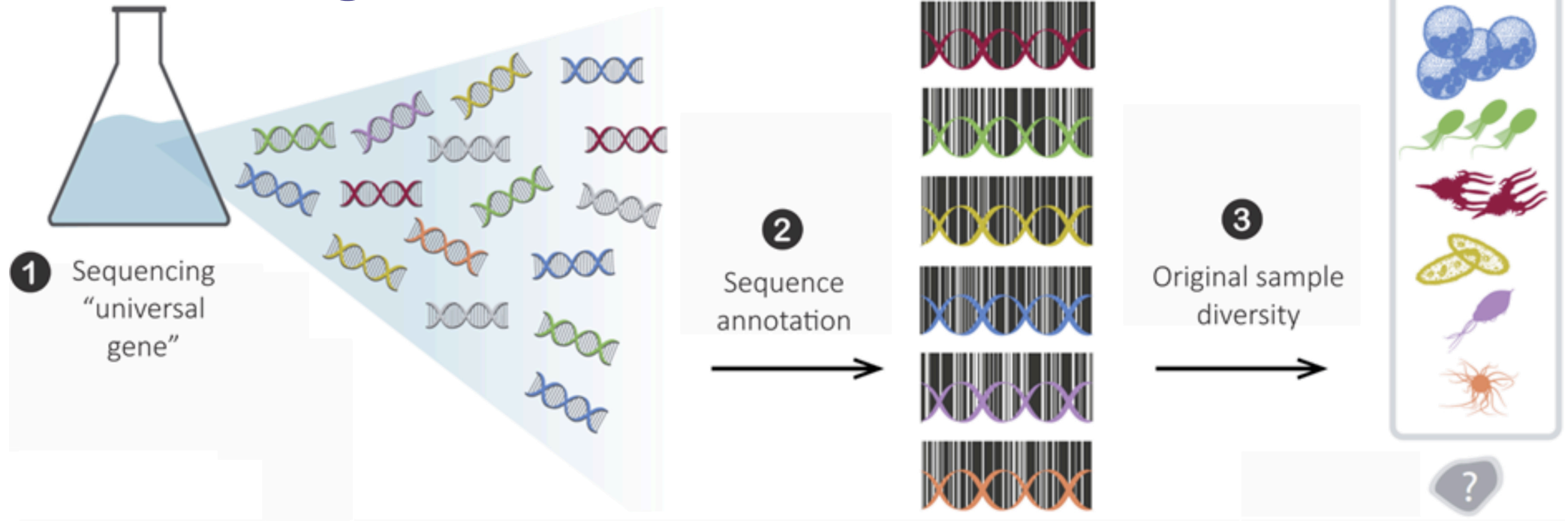
Do you want to share your work with other scientists?

Do you want to increase the visibility of your research?

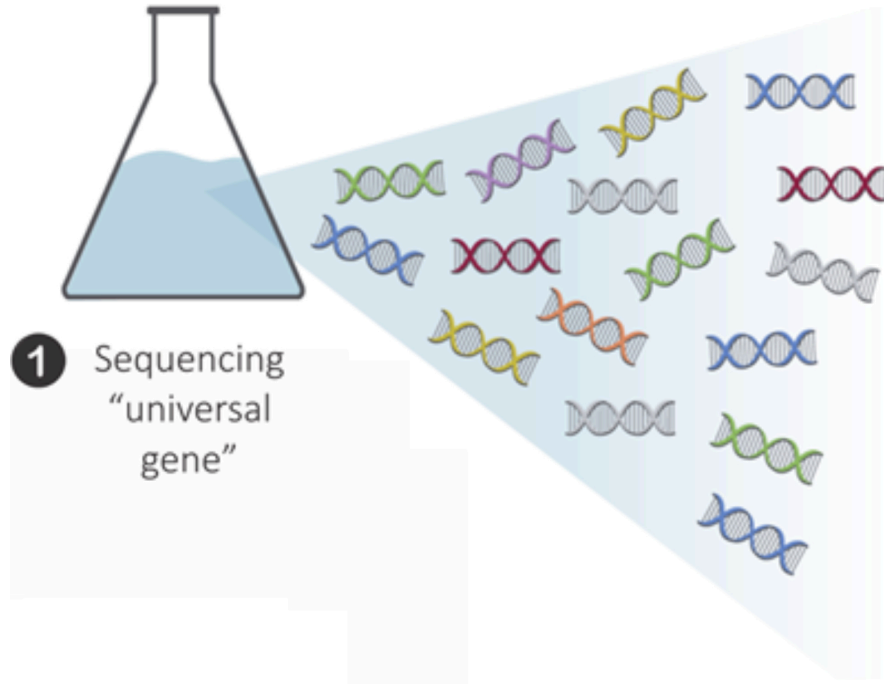
Outline

- The PR² ecosystem
- Protist Ribosomal Reference (PR²) database
- PR² primers
- metaPR²

Metabarcoding



Metabarcoding

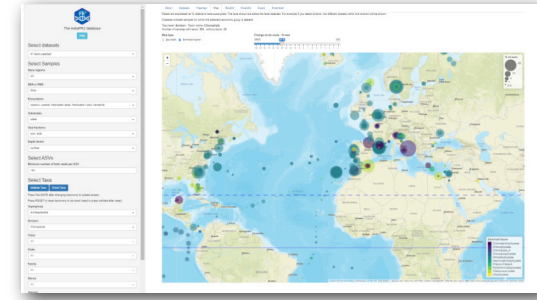
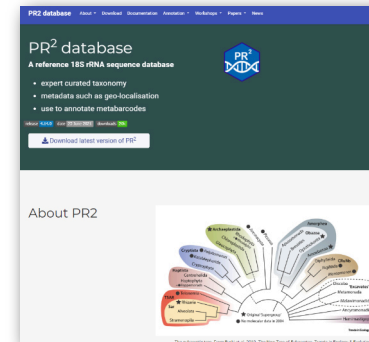


pr2-primers



pr2-database

metapr2

[illegible]

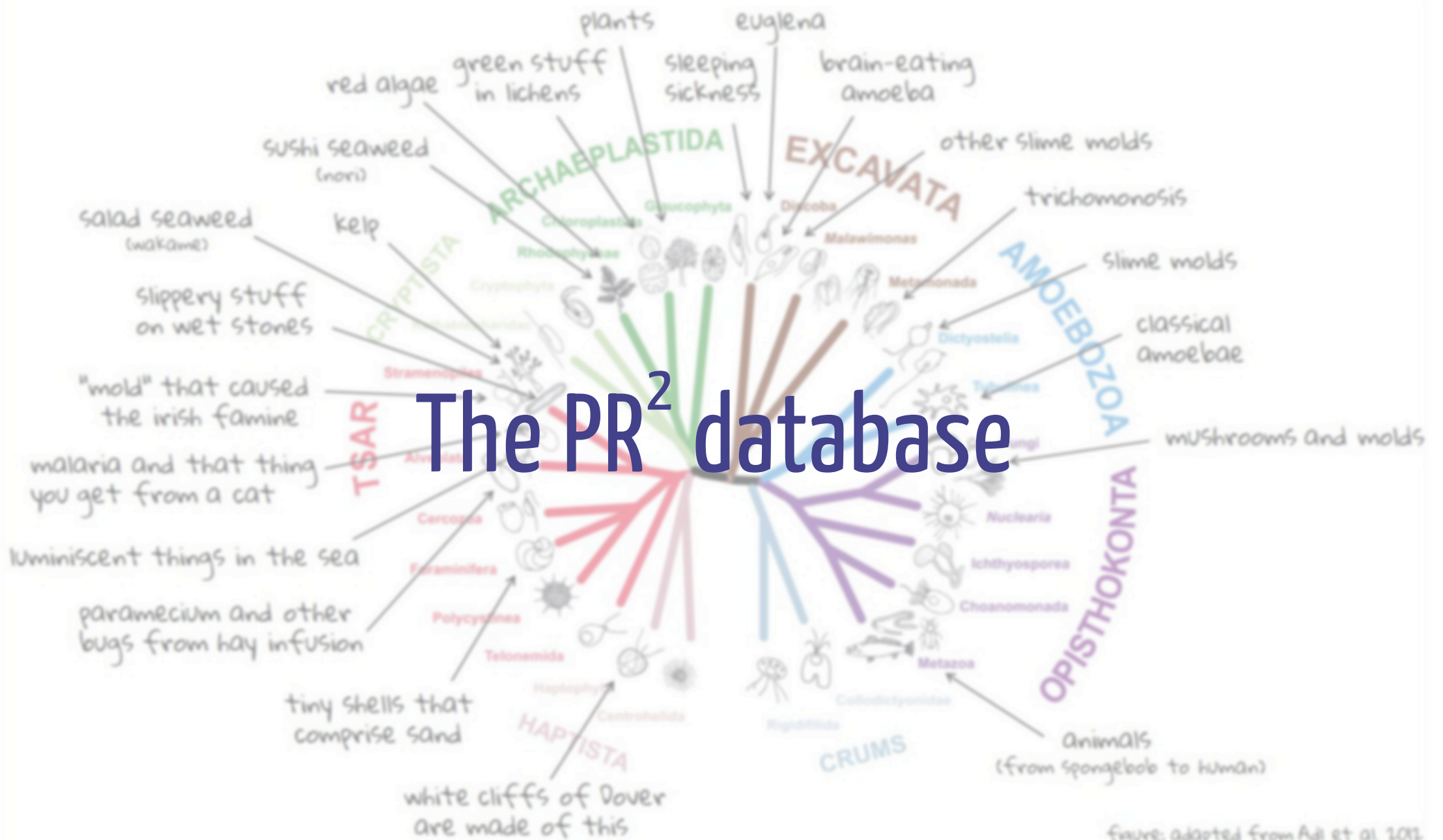


figure: adapted from Aili et al. 2012
text: @euglenaria

PR² reference database

- Created in 2010 (Biomarkers project)
- 240 199 annotated sequences
 - nuclear 18S rRNA
 - plastid 16S rRNA (PhytoRef)
 - bacteria and archaea 16S rRNA
 - rRNA operons (web interface)
- Unified taxonomy (9 ranks from domain to species)
- Quality control (e.g. > 500 bp., N < 20, no "NN")
- Metadata (e.g. coordinates, environment)
- Link to other databases (e.g. GBIF)
- Species functional annotation (e.g. ecological function)
- Web interface / Flat file
- Main use: taxonomic annotation of metabarcodes

PR² reference sequence database

- Current version : 5.0.1
- Last update : 15 May 2023
- Web interface - <https://app.pr2-database.org>
- DOI: [10.5281/zenodo.7805244](https://doi.org/10.5281/zenodo.7805244)
- Download files: <https://github.com/pr2database/pr2database/releases>

taxid	domain	supergroup	division	subdivision	class	order	family
1	Eukaryota	TSAR	Alveolata	Alveolata_X	Alveolata_XX	Alveolata_XXX	Alveolata_XXXX
2	Eukaryota	TSAR	Alveolata	Alveolata_X	Ellobiosididae	Ellobiosididae_X	Ellobiosididae_XX
3	Eukaryota	TSAR	Alveolata	Alveolata_X	Ellobiosididae	Ellobiosididae_X	Ellobiosididae_XX
4	Eukaryota	TSAR	Alveolata	Alveolata_X	Ellobiosididae	Ellobiosididae_X	Ellobiosididae_XX
5	Eukaryota	Amoebozoa	Excavata	Eumycetozoa	Myxogastria, Loriciferia	Trichidae	Demospongiae
154	Eukaryota	Obazoa	Opisthokonta	Metazoa	Porifera	Porifera_X	Porifera_XX
304	Eukaryota	Obazoa	Opisthokonta	Metazoa	Artropoda	Artropoda_X	Artropoda_XX
305	Eukaryota	TSAR	Alveolata	Alveolata_X	Alveolata_XX	Alveolata_XXX	Alveolata_XXXX
403	Eukaryota	TSAR	Rhizaria	Foramsintra	Globobulimina	Rotalia	Ammonia
474	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
475	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
476	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
477	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
478	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
479	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
480	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
481	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
482	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
483	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
484	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
485	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
486	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
487	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
488	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
489	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
490	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
491	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
492	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
493	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
494	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
495	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
496	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
497	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
498	Eukaryota	TSAR	Alveolata	Alveolata_X	Amphipoda	Amphipoda_X	Amphipoda_XX
503	Eukaryota	TSAR	Alveolata	Alveolata_X	CONFthreeP	CONFthreeP_X	CONFthreeP_XX
514	Eukaryota	TSAR	Alveolata	Alveolata_X	Colpoda	Colpoda_X	Colpoda_XX
515	Eukaryota	TSAR	Alveolata	Alveolata_X	Colpoda	Colpoda_X	Colpoda_XX
517	Eukaryota	TSAR	Alveolata	Alveolata_X	Colpoda	Colpoda_X	Colpoda_XX
519	Eukaryota	TSAR	Alveolata	Alveolata_X	Colpoda	Colpoda_X	Colpoda_XX

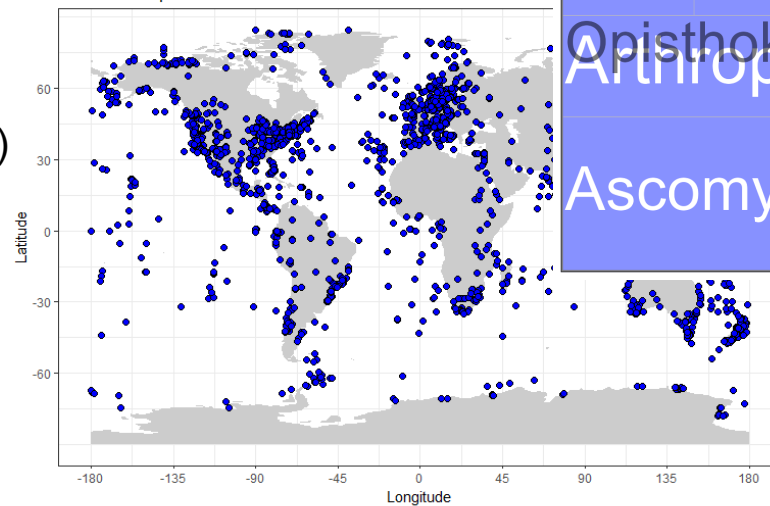
Published online 27 November 2012

Nucleic Acids Research, 2013, Vol. 41, Database issue D597–D604
doi:10.1093/nar/gks1160

The Protist Ribosomal Reference database (PR²): a catalog of unicellular eukaryote Small Sub-Unit rRNA sequences with curated taxonomy

Laure Guillou^{1,2,*}, Dipankar Bachar^{3,4}, Stéphane Audic^{1,2}, David Bass⁵, Cédric Berny⁶, Lucie Bittner^{1,2}, Christophe Boute^{1,2}, Gaëtan Burgaud⁶, Colomban de Vargas^{1,2}, Johan Decelle^{1,2}, Javier del Campo⁷, John R. Dolan⁸, Micah Dunthorn⁹, Bente Edvardsen¹⁰, Maria Holzmah¹¹, Wiebe H.C.F. Kooistra¹², Enrique Lara¹³, Noan Le Bescot^{1,2}, Ramiro Logares¹, Frédéric Mahé^{1,2}, Ramon Massana¹, Marina Montresor¹², Raphael Morard^{1,2}, Fabrice Not^{1,2}, Jan Pawlowski¹¹, Ian Probert^{14,15}, Anne-Laure Sauvadet^{1,2}, Raffaele Siano¹⁶, Thorsten Stoeck⁹, Daniel Vaulot^{1,2}, Pascal Zimmermann¹⁷ and Richard Christen^{3,4,*}

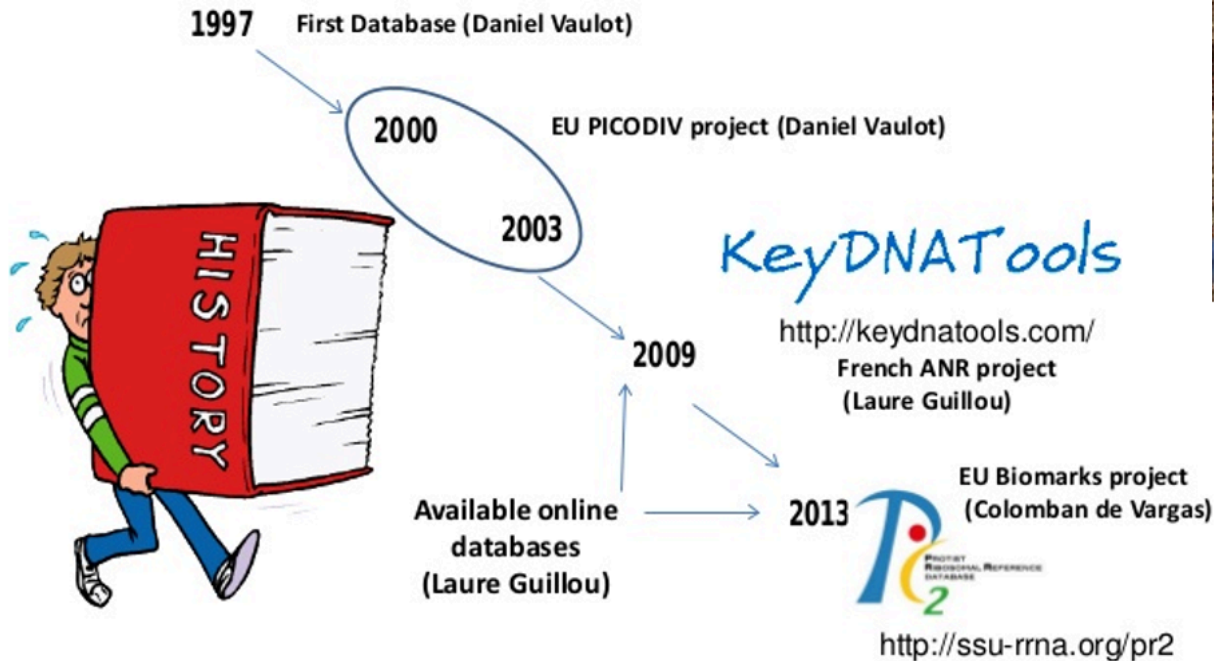
PR² - all sequences



History



- Item



Published online 27 November 2012

Nucleic Acids Research, 2013, Vol. 41, Database issue D597–D604
 doi:10.1093/nar/gks1160

The Protist Ribosomal Reference database (PR²): a catalog of unicellular eukaryote Small Sub-Unit rRNA sequences with curated taxonomy

Laure Guillou^{1,2,*}, Dipankar Bachar^{3,4}, Stéphane Audic^{1,2}, David Bass⁵, Cédric Berney⁵, Lucie Bittner^{1,2}, Christophe Boutte^{1,2}, Gaétan Burgaud⁶, Colomban de Vargas^{1,2}, Johan Decelle^{1,2}, Javier del Campo⁷, John R. Dolan⁸, Micah Dunthorn⁹, Bente Edvardsen¹⁰, Maria Holzmann¹¹, Wiebe H.C.F. Kooistra¹², Enrique Lara¹³, Noan Le Bescot^{1,2}, Ramiro Logares⁷, Frédéric Mahé^{1,2}, Ramon Massana⁷, Marina Montresor¹², Raphael Morard^{1,2}, Fabrice Not^{1,2}, Jan Pawlowski¹¹, Ian Probert^{14,15}, Anne-Laure Sauvadet^{1,2}, Raffaele Siano¹⁶, Thorsten Stoeck⁹, Daniel Vaultot^{1,2}, Pascal Zimmermann¹⁷ and Richard Christen^{3,4,*}

Annotation - Eukref

Workshop Roscoff 2018

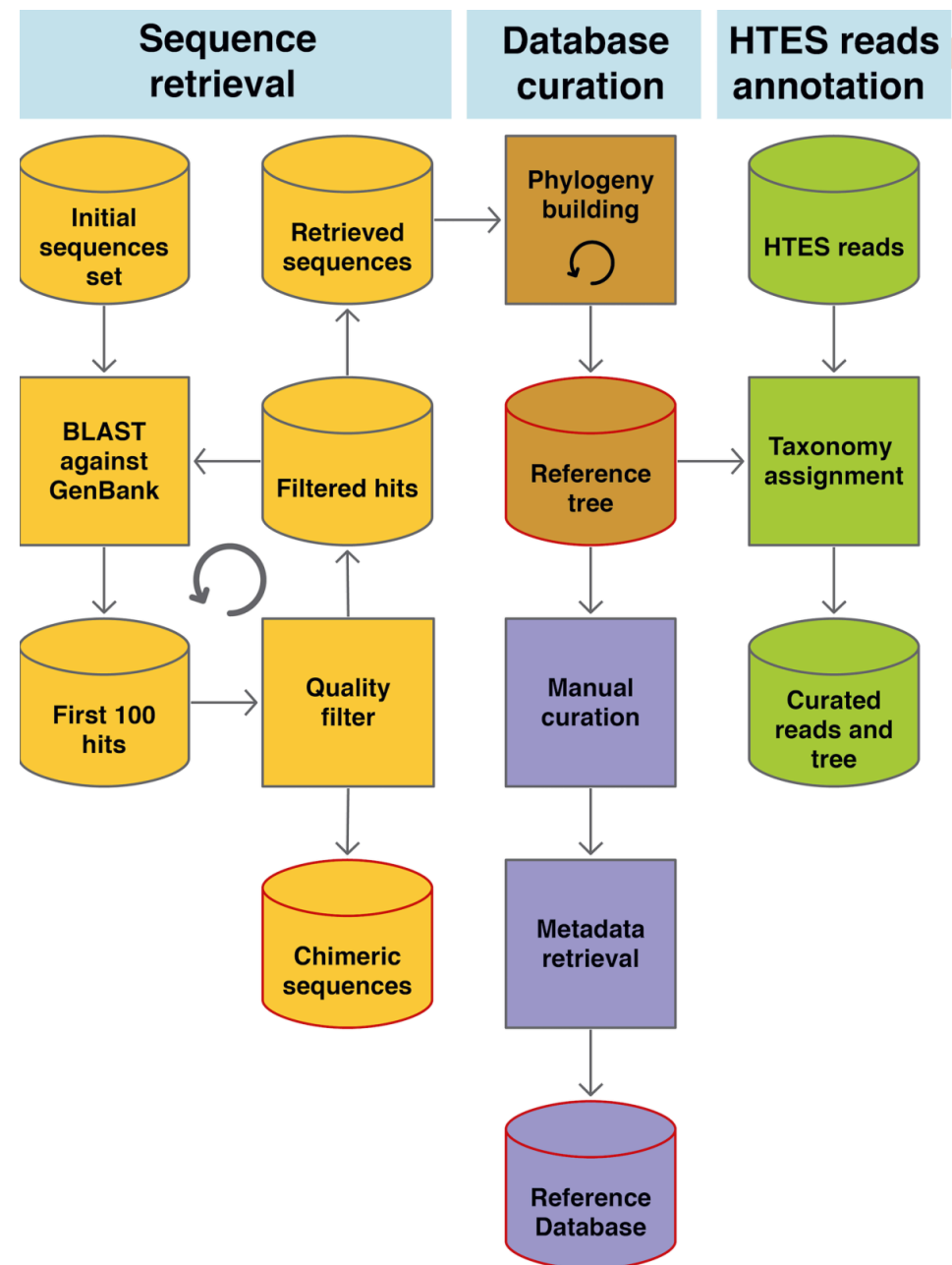
Javier del Campo

Last updated on 2019-11-29 ·  **eukref**



Our third workshop took place at the **Station Biologique Roscoff** in **Roscoff**, France from **November 5 to 9 2018**. This effort brought together taxonomists with expertise in individual lineages that span the eukaryotic tree of life to curate reference 18S rDNA sequences of these lineages by incorporating knowledge of phylogenetic, morphological, and/or environmental contextual data. **This third edition was particularly (but not exclusively) focused on Diatoms, Dinoflagellates, and Green Algae.** During the workshop,

<https://pr2-database.org/eukref/about/>



Annotation - Eukref



- Simplified pipeline
- Adapt more closely to PR2 structure
- Groups annotated
 - Picozoa
 - Chloropicophyceae
 - Red algae
 - Metazoa
 - Dinoflagellates
 - Basal fungi
 - ...

<https://pr2-database.org/eukref/about/>

Annotation Workshop Barcelona 2023

Javier del Campo

Last updated on 2023-06-28 · eukref



Our fourth EukRef workshop took place at the [Institute of Marine Sciences](#) in [Barcelona](#), Spain from **July 3 to July 7 2023**.

The workshop was made possible thanks to the generous contribution of the [Moore Foundation](#).

Organizers

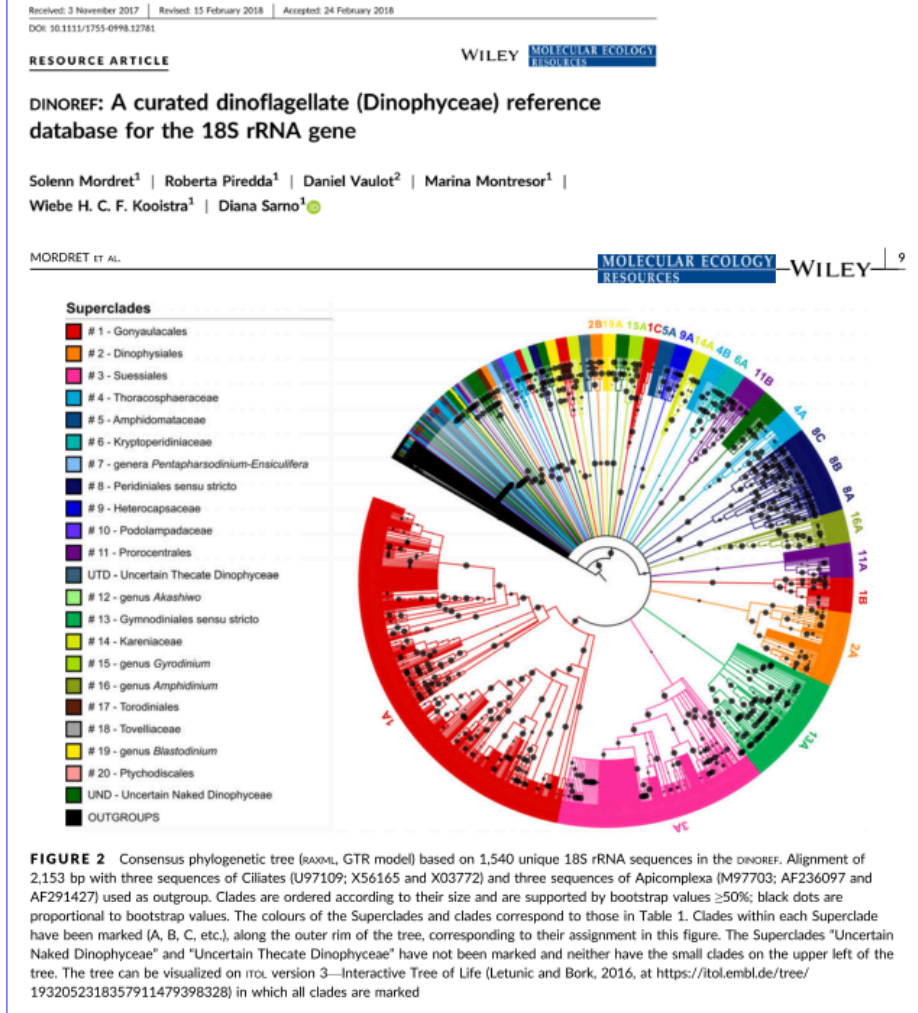
- Javier del Campo, Barcelona
- Daniel Vaultot, CNRS Roscoff
- Chris Lane, University of Rhode Island

Annotation - Contributions



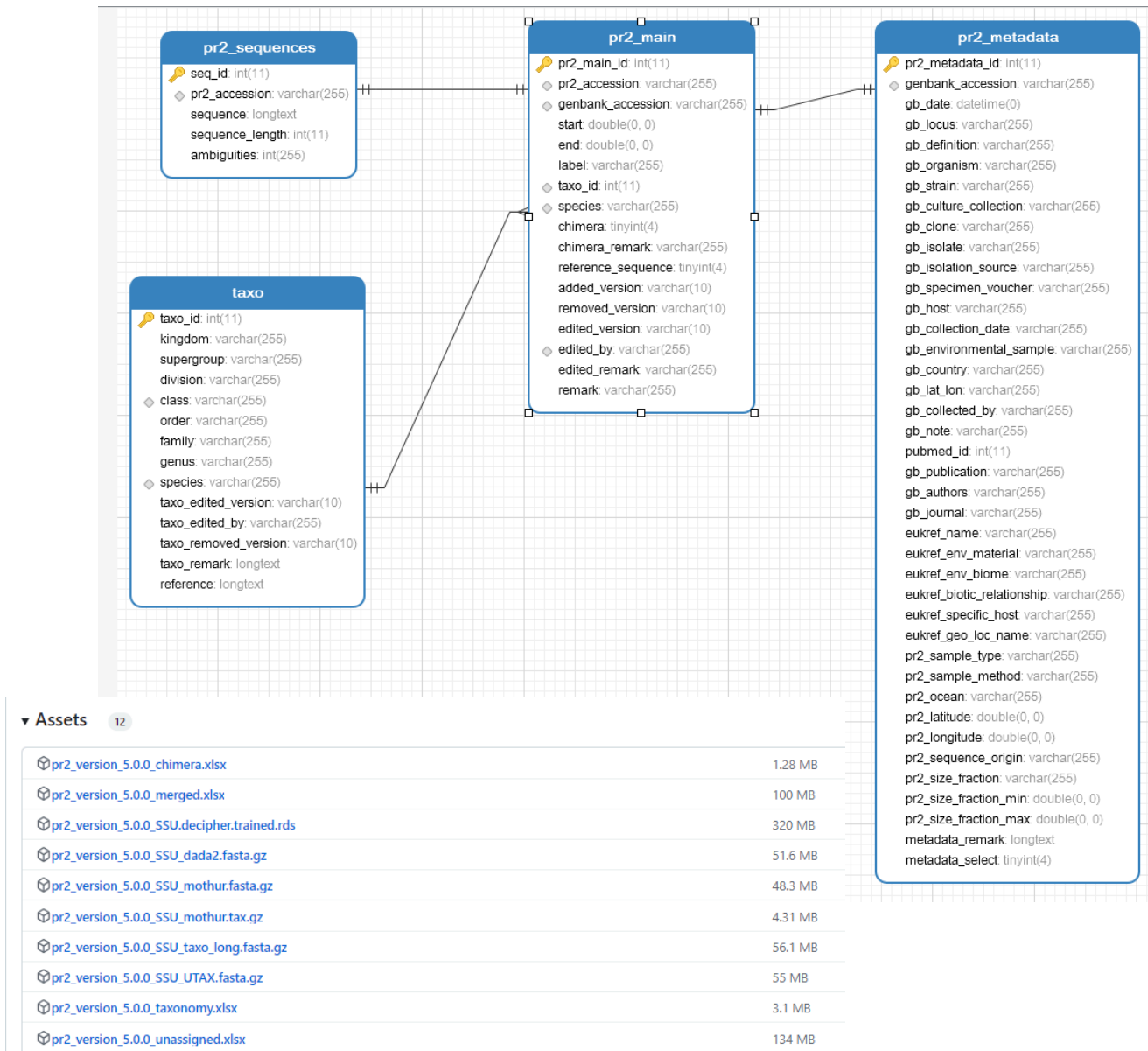
Groups with updated annotations

Division	Class	Group	Who	Date	Version
Alveolata	Apicomplexa		J. del Campo	2019	4.12, 4.14
Alveolata	Ciliates		W. Ting, C. Bachy	2017	4.7
Alveolata	Ciliates		V. Boscaro, L. Santoferrara, E. Gentekaki and Q. Zhang	2018	4.11
Alveolata	Dinoflagellates		S. Mordret, D. Sarno	2018	4.9
Alveolata	Dinoflagellates	Suessiales	J. del Campo	2021	4.13
Chlorophyta			M. Tragin, A. Lopes dos Santos	2015	3.0
Excavata			J. del Campo	2021	4.14
Haptophyta			B. Edvardsen	2015	4.0
Prasinodermophyta			D. Vaultot	2021	4.13
Rhizaria	Collodaria		T. Biard	2015	2.0
Rhizaria	Foraminifera		R. Morard	2021	
Rhizaria	Radiolaria		M. Mendez Sandin	2021	
Stramenopiles			R. Massana	2019	4.12
Stramenopiles		Cafeteria	Alex Schoenle	2021	4.14
Stramenopiles	Labyrinthulomycetes		J. del Campo	2021	4.14
Stramenopiles	Diatoms	Chaetoceros	C. Gaonkar	2019	4.12
Stramenopiles	Diatoms	Thalassiosirales	L. Arsenieff	2021	4.13
Stramenopiles	Bolidophyceae		D. Vaultot	2017	4.6
Stramenopiles	Pelagophyceae		A.M. Cabello	2021	4.13
Stramenopiles	Chrysophyceae		D. Vaultot	2021	4.13



Management

- MySQL database
- R scripts for:
 - importing
 - exporting
 - validating
- Data provided as
 - text files (for dada2, mothur)
 - fasta (phylogeny)
 - R package



pr2database 4.14.0

Get startedReferenceArticlesChangelog

Get started

Daniel Vaultot

The PR2 database is provided as a R package called **pr2database**. This page provides instruction to install and use the package.

Installation

Install from the GitHub web site using the devtools package

```
install.packages(devtools)
devtools::install_github("pr2database/pr2database")
```

```
* installing *source* package 'pr2database' ...
** R
** data
*** moving datasets to lazyload DB
** byte-compile and prepare package for lazy loading
** help
*** installing help indices
    converting help for package 'pr2database'
      finding HTML links ... fini
      pr2                                html
** building package indices
** testing if installed package can be loaded
*** arch - i386
*** arch - x64
* DONE (pr2database)
In R CMD INSTALL
```

The R pr2database package

Daniel Vaultot edited this page 4 days ago · 7 revisions

The PR2 database is now provided as a R package

Installation

Install from the GitHub web site using the devtools package

```
install.packages(devtools)
devtools::install_github("vaulot/pr2database")
```

Selecting sequences from a specific taxon

Let us select all the available sequences for the Mamiellophyceae *Ostreococcus*

```
# Filter only the sequences for which the column genus contains Ostreococcus
pr2_ostreo <- pr2 %>% dplyr::filter(genus == "Ostreococcus")

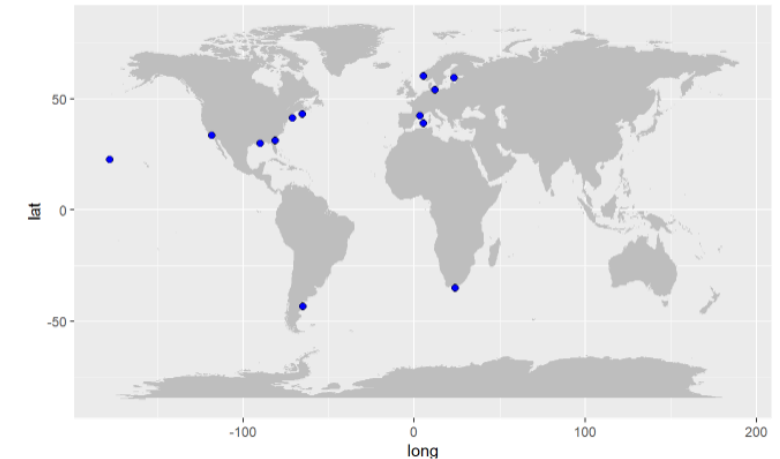
# Select only the columns of interest
pr2_ostreo <- pr2_ostreo %>% dplyr::select( genbank_accession, species,
                                           pr2_sample_type, gb_strain, gb_clone,
                                           pr2_latitude, pr2_longitude,
                                           sequence_length, sequence )
```

Drawing a map of sequence locations

```
library(maps)
world <- map_data("world")

ggplot() +
  geom_polygon(data = world, aes(x=long, y = lat, group = group), fill="grey") +
  coord_fixed(1.3) +
  geom_point(data=pr2_ostreo, aes(x=pr2_longitude, y=pr2_latitude), fill="blue", size=2, shape=21) +
  ggtitle("Ostreococcus")
```

Ostreococcus



Web interface

<https://app.pr2-database.org>



- Built with R shiny

- Panels

- ☐ Taxonomy
- ☐ Sequences
- ☐ Download
 - ☐ selected
 - ☐ full
- ☐ Query



PR2 18S rRNA database v. 5.0.1
221,085 sequences

Help Disconnect

Sample type
culture, environmental, isolate, specimen, unknown

Minimum sequence length
374 2,000

Select Taxa
Validate Taxa Reset Taxa
VALIDATE after changes to update screen.
RESET + VALIDATE to cancel choices

Domain
Eukaryota

Supergroup
TSAR

Division
Alveolata

Subdivision
Dinoflagellata

Class
Dinophyceae

Order
All

Taxonomy Sequences Download selected sequences Download full database Query About

Taxonomy table
[Information about database structure](#)
Species with valid WoRMS database entries display link to WoRMS.

Press **Validate Taxa** to load or update taxonomy table.
Taxo level: class - Taxon name: Dinophyceae - Number of species: 616
Show 50 entries

domain	supergroup	division	subdivision	class	order	family	genus	species	n_sequences	mixoplankton	worms_id
Eukaryota	TSAR	Alveolata	Dinoflagellata	Dinophyceae	Dinophyceae_X	Dinophyceae_XX	Abedinium	Abedinium_dasypus	3		233177
Eukaryota	TSAR	Alveolata	Dinoflagellata	Dinophyceae	Dinophyceae_X	Dinophyceae_XX	Alladinium	Alladinium_reticulatum	2		841392
Eukaryota	TSAR	Alveolata	Dinoflagellata	Dinophyceae	Dinophyceae_X	Dinophyceae_XX	Amphidiniella	Amphidiniella_sedentaria	2	CM	233845
Eukaryota	TSAR	Alveolata	Dinoflagellata	Dinophyceae	Dinophyceae_X	Dinophyceae_XX	Cucumeridinium				
Eukaryota	TSAR	Alveolata	Dinoflagellata	Dinophyceae	Dinophyceae_X	Dinophyceae_XX	Cucumeridinium				
Eukaryota	TSAR	Alveolata	Dinoflagellata	Dinophyceae	Dinophyceae_X	Dinophyceae_XX	Cucumeridinium				
Eukaryota	TSAR	Alveolata	Dinoflagellata	Dinophyceae	Dinophyceae_X	Dinophyceae_XX	Dapsilidinium				
Eukaryota	TSAR	Alveolata	Dinoflagellata	Dinophyceae	Dinophyceae_X	Dinophyceae_XX	Dinophyceae				
Eukaryota	TSAR	Alveolata	Dinoflagellata	Dinophyceae	Dinophyceae_X	Dinophyceae_XX	Fensomea				
Eukaryota	TSAR	Alveolata	Dinoflagellata	Dinophyceae	Dinophyceae_X	Dinophyceae_XX	Glenaulax				
Eukaryota	TSAR	Alveolata	Dinoflagellata	Dinophyceae	Dinophyceae_X	Dinophyceae_XX	Halostylodinium				
Eukaryota	TSAR	Alveolata	Dinoflagellata	Dinophyceae	Dinophyceae_X	Dinophyceae_XX	Madanidinium				
Eukaryota	TSAR	Alveolata	Dinoflagellata	Dinophyceae	Dinophyceae_X	Dinophyceae_XX	Pileidinium				
Eukaryota	TSAR	Alveolata	Dinoflagellata	Dinophyceae	Dinophyceae_X	Dinophyceae_XX	Oodiniaceae	Oodinium			
Eukaryota	TSAR	Alveolata	Dinoflagellata	Dinophyceae	Dinophyceae_X	Phytodiniaceae	Stylodinium				
Eukaryota	TSAR	Alveolata	Dinoflagellata	Dinophyceae	Dinophyceae_X	Ptychodiscaceae	Ptychodiscus				
Eukaryota	TSAR	Alveolata	Dinoflagellata	Dinophyceae	Dinophyceae_X	Tovelliacae	Esotrodinium				
Eukaryota	TSAR	Alveolata	Dinoflagellata	Dinophyceae	Dinophyceae_X	Tovelliacae	Jadwisia				

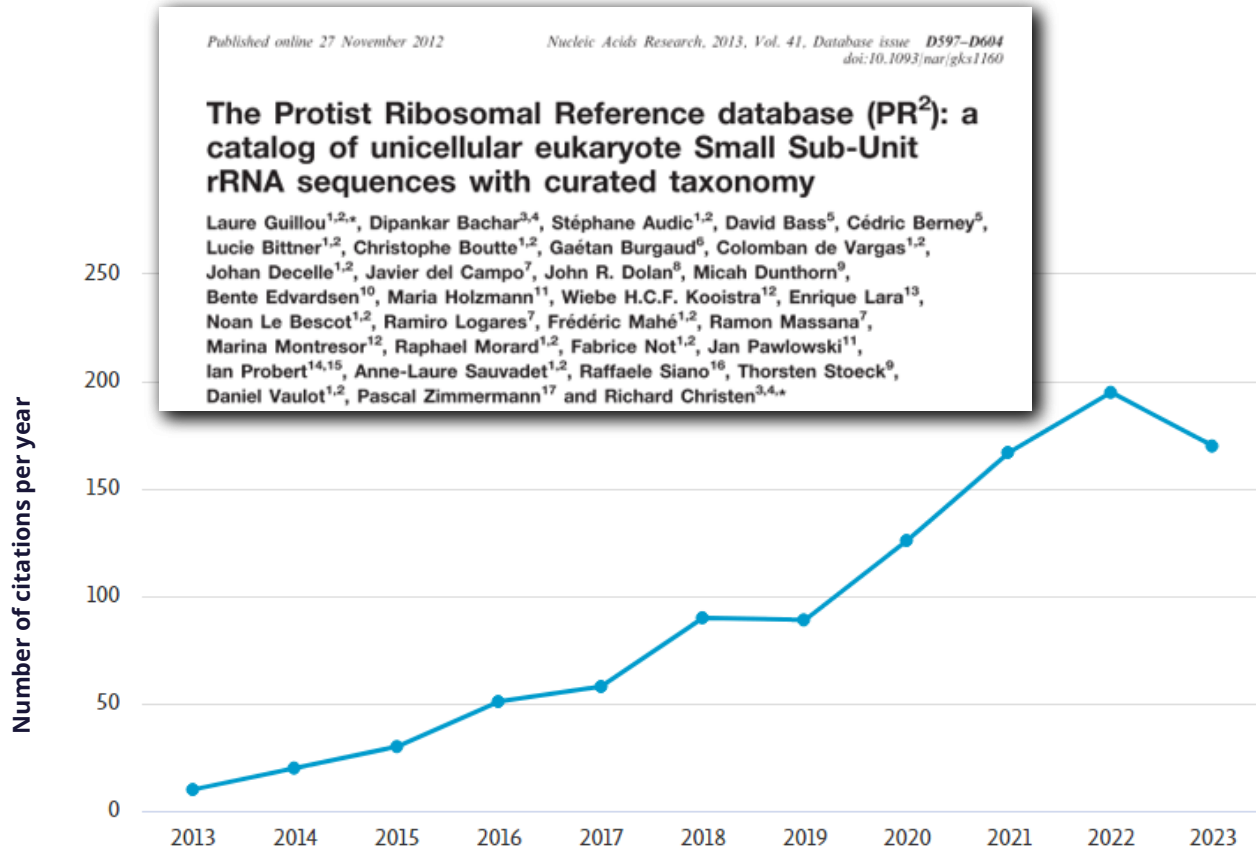
Taxonomy Sequences Download selected sequences Download full database Query About

BLAST-like search for PR2 sequences similar to query sequence.
All PR2 sequences are searched but only those above % identity min are shown
% identity min
80 82 84 86 88 90 92 94 96 98 100
Query - at least 130 bp
>KU244632.1 Micromonas commoda 18S ribosomal RNA gene, partial sequence
TGTTCTAAGTATTAAGCGTTTACTGCTGAACCTGCGATGCTTCAATCAACGCAATAGTTCTTTGGTG
TGTTTACTACATGGATAACCGTAGTAATCTAGAGCTAATACATGCTAATCCGACCTTCGGAAGGGAG
GTATTATTAGATAAAGACCGACCTGCTGCTGCGGTGAATCATGATAACTTCACGACCGCATGGCTGT
CGCGCGCGGTGTTCCATTCAATTTCTGCGCTATCAACTTTCGACGATGATAGAGGCTACCGTGATG
Search - be patient

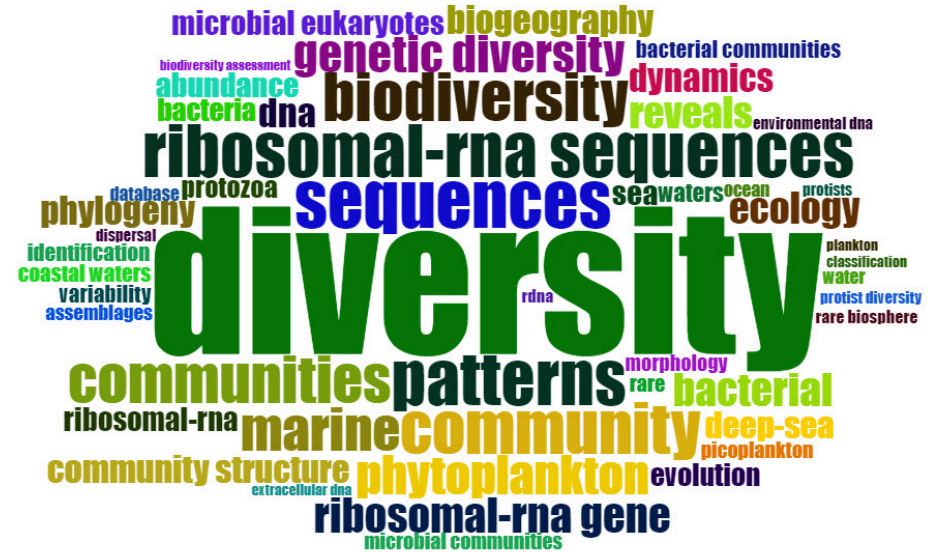
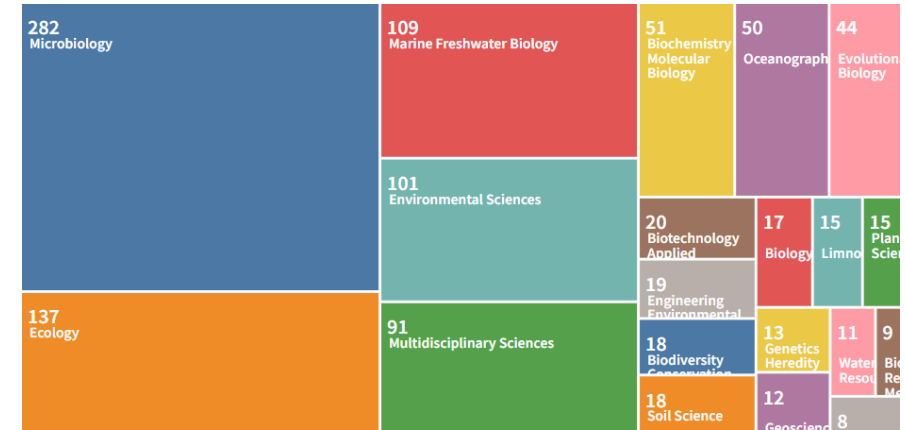
Matching PR2 sequences
Download filtered results (zip)
Show 50 entries

pr2_accession	species	pid	mismatches	gaps	query_start	query_end	target_start	target_end
KU244663.1.1703_U	Micromonas_pusilla	99.8	4	0	1	1703	1	1703
KU743492.1.1777_U	Micromonas_commoda_A1	99.8	4	0	1	1703	55	1757
AY955011.1.1727_U	Micromonas_commoda_A1	99.8	4	0	1	1703	1	1703
AY955002.1.1727_U	Micromonas_commoda_A1	99.8	4	0	1	1703	1	1703
HM191693.1.1777_U	Micromonas_commoda_A1	99.8	4	0	1	1703	55	1757
CP001575.879464.881247_U	Micromonas_commoda_A1	99.8	4	0	1	1703	56	1756
JX188376.1.1738_U	Micromonas_commoda_A1	99.8	4	0	1	1703	36	1738
CP001575.894647.896426_U	Micromonas_commoda_A1	99.8	4	0	1	1703	55	1757
AB183589.1.1734_U	Micromonas_commoda_A1	99.8	4	0	1	1699	36	1734
AY955001.1.1727_U	Micromonas_commoda_A1	99.6	7	0	1	1703	1	1703
KF129910.1.1738_U	Micromonas_commoda_A1	99.6	7	0	1	1703	36	1738
AY955006.1.1727_U	Micromonas_commoda_A1	99.6	7	0	1	1703	1	1703
AY955007.1.1727_U	Micromonas_commoda_A1	99.6	7	0	1	1703	1	1703
KF129834.1.1738_U	Micromonas_commoda_A1	99.6	6	0	1	1703	36	1738

More than 1,700 papers citing PR²



 As of January/February 2022, this [highly cited paper](#) received enough citations to place it in the top 1% of the academic field of **Biology & Biochemistry** based on a highly cited threshold for the field and publication year.



Latest version - 5.1 - April 2025

- Update of some groups
 - Picozoa
 - Choropicophyceae
 - Rhodophyta
 - Dinophyceae
 - Foraminifera
- New sequences from GenBank
- Fixing taxonomy
- Link to GBIF
- Functional traits
 - HAB (Harmful Algal Bloom) species
 - ecological function - from Sommeria-Klein et al. 2021
- Web interface to Ribosomal Operon Database (ROD)

PR2 18S rRNA database v. 5.1.0
Ribosomal Operon database v. 1.2.0
309,663 sequences

Genes
operon_rRNA

☐ Only reference sequences

Sample type
culture, environmental, isolate, specimen, unknown

Minimum sequence length
0 750 1,500 2,250 3,000 3,750 4,500 5,250 6,000

☐ Only sequences from Harmful algal blooms (HAB) species

☐ Only sequences from Mixoplankton species

Select Taxa
Validate Taxa Reset Taxa

VALIDATE after changes to update screen.
RESET + VALIDATE to cancel choices

Domain
Eukaryota

Supergroup
Archaeplastida

Taxonomy Sequences Download selected sequences Download full database Query About

Sequence table
Information about database structure

Press **Validate Taxa** to update taxonomy and load sequence table.
You can download these sequences and the corresponding metadata in the "Download selected sequences" tab.

Taxo level: **supergroup** - Taxon name: **Archaeplastida** - Number of sequences: **47975**

Show **50** entries

accession_genbank_link	gene	species	gb_organism	sequence_length	pr2_sample_type	reference_sequence
GCA_026401115JAPMLY01000537.1/97891-103568	operon_rRNA	Tetraselmis_chuii	Tetraselmis chuii	5678	culture	Yes
GCA_022414725JCM039462.1/194499987-194493578	operon_rRNA	Microglona_sp.	Microglona sp. YARC	6410	isolate	Yes
GCA_022414725JCM039463.1/33960276-33966809	operon_rRNA	Microglona_sp.	Microglona sp. YARC	6534	isolate	
GCA_022414725JAJSRW010000615.1/25524-32056	operon_rRNA	Microglona_sp.	Microglona sp. YARC	6533	isolate	
GCA_022414725JAJSRW010000654.1/9180-15799	operon_rRNA	Microglona_sp.	Microglona sp. YARC	6620	isolate	
GCA_022414725JAJSRW010002046.1/3467-9939	operon_rRNA	Microglona_sp.	Microglona sp. YARC	6473	isolate	
GCA_016834605JAEHOC010000025.1/1459517-1465257	operon_rRNA	Chlamydomonas_incerta	Chlamydomonas incerta	5741	culture	Yes
GCA_000002595JCM008969.1/5027808-5033549	operon_rRNA	Chlamydomonas_reinhardtii	Chlamydomonas reinhardtii	5742	culture	Yes
GCA_013389655JCM023806.1/6844-1106	operon_rRNA	Chlamydomonas_reinhardtii	Chlamydomonas reinhardtii	5739	culture	Yes
GCA_013389655JCM023819.1/4222930-4228634	operon_rRNA	Chlamydomonas_reinhardtii	Chlamydomonas reinhardtii	5705	culture	
GCA_018267845JCM031089.1/7090-1349	operon_rRNA	Chlamydomonas_reinhardtii	Chlamydomonas reinhardtii	5742	culture	Yes

What's next for **pr2-database**

- Integration of taxonomy into GBIF (in progress)
- Need more participation from taxonomy experts
- Full operon
 - Include metagenomic data ?
- Functional annotation
 - Trophic mode (photo, hetero, mixo...)
 - Size group (pico, nano, micro...)
- Other genes: *coi*

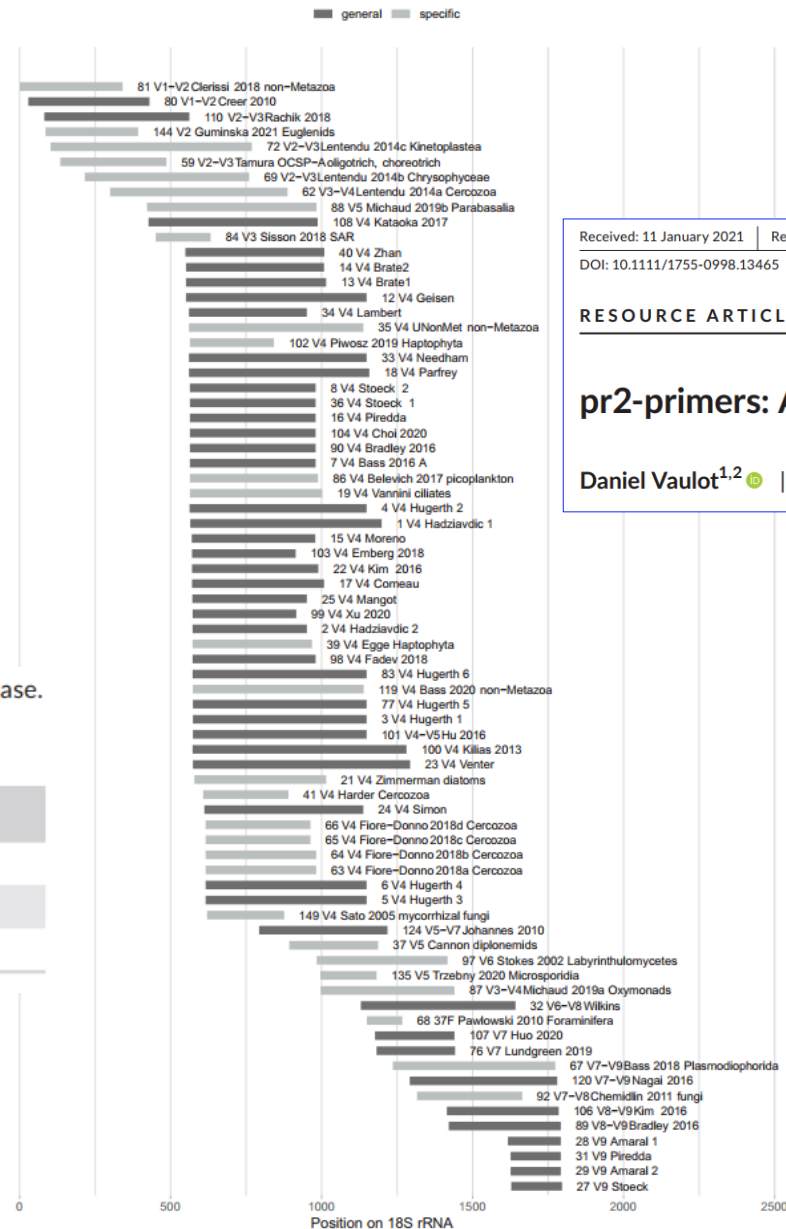
The PR² primer database

18S rRNA primers

- Wide diversity of primers and sets
- No database for protists
- Taxonomic specificity of primers?

TABLE 1 Summary of primers listed in the pr2-primers database. General primers target all eukaryotes and specific primers only certain taxonomic groups

Direction	General primers	Specific primers
fwd	55	89
rev	53	88
Total	108	177



Received: 11 January 2021 | Revised: 31 May 2021 | Accepted: 6 July 2021

DOI: 10.1111/1755-0998.13465

MOLECULAR ECOLOGY
RESOURCES WILEY

RESOURCE ARTICLE

pr2-primers: An 18S rRNA primer database for protists

Daniel Vaultot^{1,2} | Stefan Geisen^{3,4,5} | Frédéric Mahé^{6,7} | David Bass^{8,9}

pr2-primers database



The PR2 primer database

Primers

column	definition
primer_id	primer id in pr2-primers database
name	original name of the primer
synonyms	synonyms found in the literature
sequence	
sequence revcomp	reverse complement of the sequence
direction	forward (fwd) or reverse (rev)
start_yeast	start of primer relative to FU970071
end_yeast	end of primer relative to FU970071
specificity	is the primer specific of a group
reference	original reference where primer was first defined
doi	link to original paper

Download primers

Columns to show:

- ☒ primer_id
- ☒ gene
- ☒ organelle
- ☒ direction
- ☒ name
- ☐ synonyms
- ☒ sequence
- ☐ sequence revcomp
- ☒ length
- ☒ start_yeast
- ☐ end_yeast
- ☒ specificity
- ☒ reference
- ☐ doi
- ☐ doi_html

About Primers Primer sets Amplification - overview Amplification - details Test your primer set

Show 25 entries

Search:

primer_id	gene	organelle	direction	name	sequence	length	start_yeast	specificity	reference
123	16S rRNA	plastid	fwd	Pla491F	GAGGAATAAGCATCGGCTAA	20		plastid	Fuller et al. (2007)
124	16S rRNA	plastid	rev	PP936R	CCTTTGAGTTTCAYYCTTGC	20		plastid	https://biomarks.eu/pp936r
212	16S rRNA	plastid	rev	OXY1313R	CTTCAYGYAGGCGAGTTGCAGC	22			West et al. (2001)
213	16S rRNA	plastid	fwd	OXY107F	GGACGGGTGAGTAACGCGTGR	21			West et al. (2001)
71	18S rRNA	nucleus	fwd	PF1	TGCGCTACCTGGTTGATCCTGCC	23	-5		Keeling, 2002
78	18S rRNA	nucleus	fwd	EukA	AACCTGGTTGATCCTGCCAGT	21	0		Medlin et al. 1988
81	18S rRNA	nucleus	fwd	Euk328F	ACCTGGTTGATCCTGCCAG	19	1		Moon et al. 2001
138	18S rRNA	nucleus	fwd	18SV1V2F	ACCTGGTTGATCCTGCCA	18	1	non-Metazoa	Clerissi et al. (2018)
220	18S rRNA	nucleus	fwd	NSF4/18	CTGGTTGATYCTGCCAGT	18	3		Hendriks et al. (1989)
168	18S rRNA	nucleus	fwd	Pbr1	GGTTGATCCTGCCAGTAGTC	20	5	Plasmodiophora	Niwa et al. 2011
169	18S rRNA	nucleus	rev	Pbr1r	GACTACTGGCAGGATCAACC	20	5	Plasmodiophora	Niwa et al. 2011
109	18S rRNA	nucleus	fwd	SF2Dark	GTTGATCCTGCCAGTAGTGT	20	6	Myxomycetes	Fiore-Donno (2016)

In silico analysis against PR²



The PR2 primer database

- Mismatches #
- Mismatches position
- Amplicon size

Precomputed results for primer sets

Against PR2 sequence database

Primer set
004 - V4 Hugerth_2 - general

Kingdom
Eukaryota

Supergroup
All

Division
All

Class
All

Update plot

About Primers Primer sets Amplification - overview Amplification - details

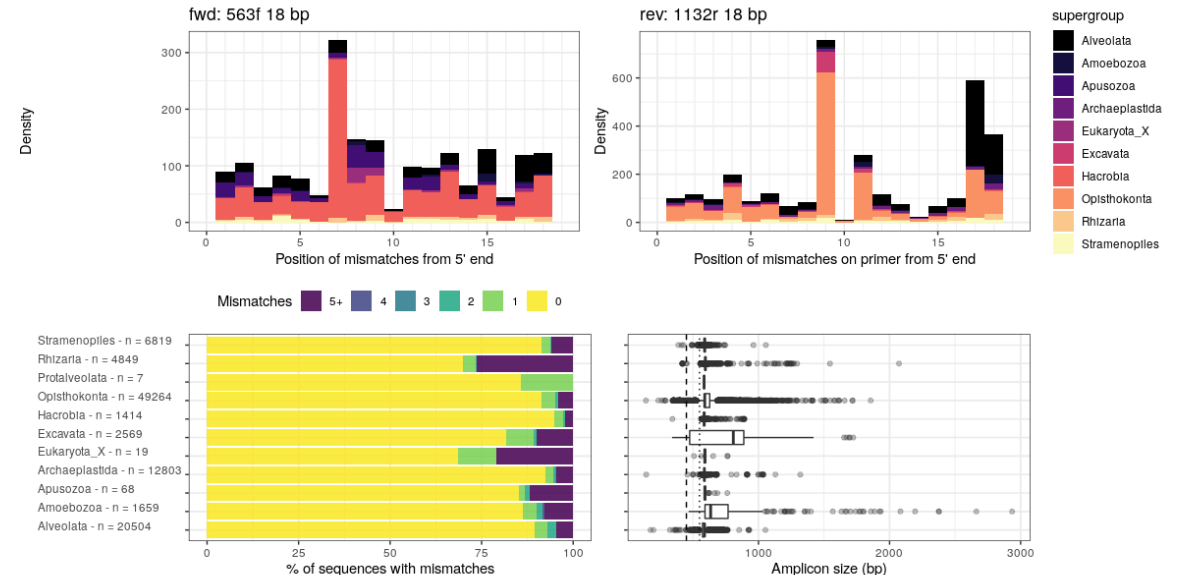
Test your primer set

Overall statistics

Archaea	% sequences matching forward primer	82.29
Archaea	% sequences matching reverse primer	100.00
Archaea	% sequences amplified	82.29
Archaea	mean amplicon size	414.77
Bacteria	% sequences matching forward primer	98.99
Bacteria	% sequences matching reverse primer	98.70
Bacteria	% sequences amplified	97.74
Bacteria	mean amplicon size	409.80
Eukaryota	% sequences matching forward primer	95.76
Eukaryota	% sequences matching reverse primer	98.29
Eukaryota	% sequences amplified	94.38
Eukaryota	mean amplicon size	604.08

Precomputed

Top panel: Location of mismatches for forward and reverse primer. Bottom panel left: number of mismatches. Bottom panel right: amplicon size



Test your own



The PR2 primer database

Test your primer/probe

Primer/Probe is tested against the PR2 database.

Use only IUPAC characters (**ACGTRY**SWKMBDHVN).

Length of primers: between 15 and 30 bp.

Primer/probe (5' -> 3')

CTTCGAGCCCCCAACTTT

Chlorophyta
probe

Max mismatches

☒ 0 ☐ 1 ☐ 2

Type

☐ primer fwd ☒ primer rev/probe

Run

Kingdom

Eukaryota

Supergroup

All

Division

All

Class

All

Update plot

About

Primers

Primer sets

Amplification - overview

Amplification - details

Test your primer/probe

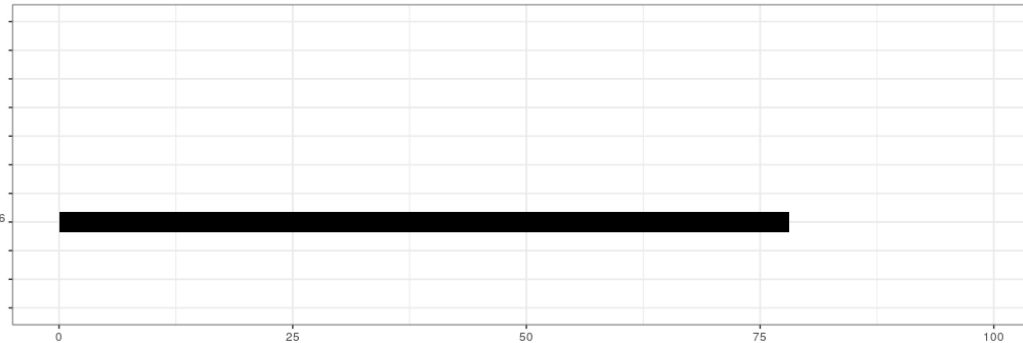
Test your primer set

Overall statistics

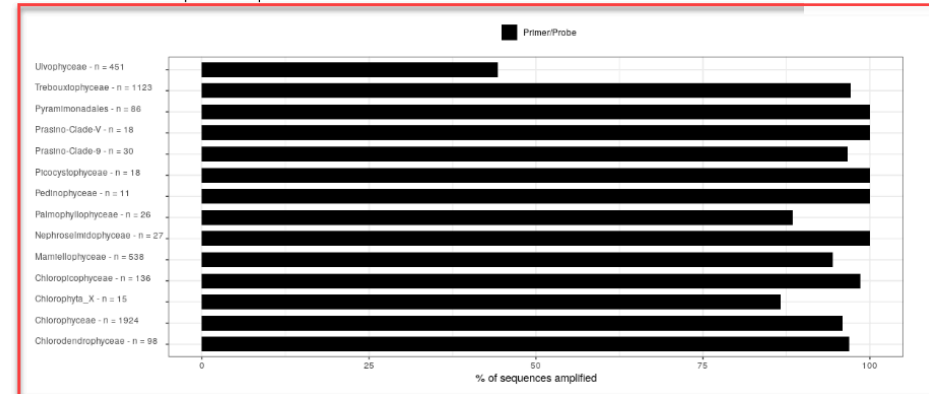
Archaea	% sequences matching primer/probe	0.00
Bacteria	% sequences matching primer/probe	0.00
Eukaryota	% sequences matching primer/probe	9.88

Download results

Stramenopiles - n = 5429
Rhizaria - n = 3308
Protalveolata - n = 7
Opisthokonta - n = 41213
Hacrobia - n = 1262
Excavata - n = 2186
Eukaryota_X - n = 16
Archaeplastida - n = 10386
Apusozoa - n = 58
Amoebozoa - n = 1360
Alveolata - n = 17059

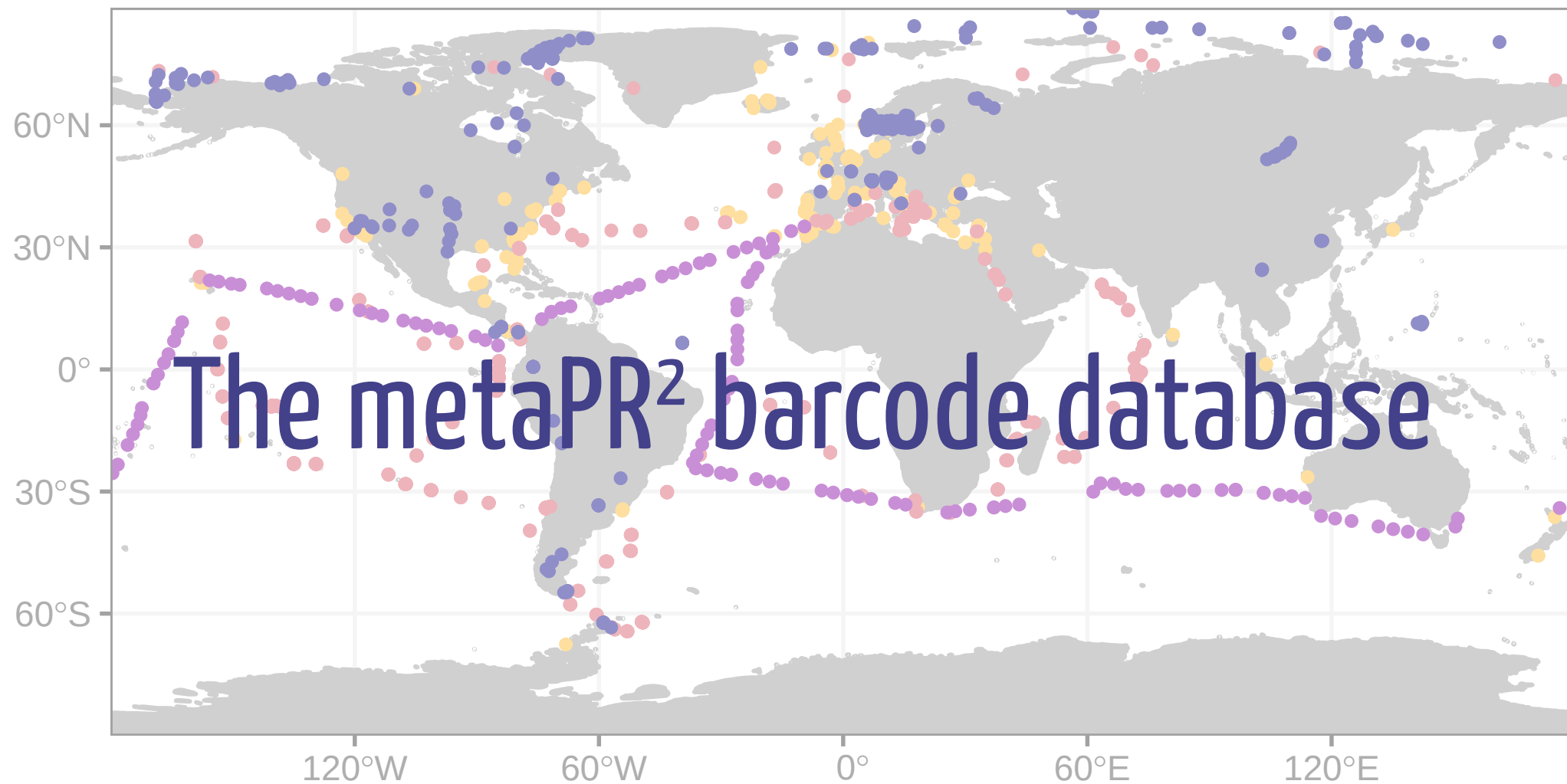


% of sequences amplified

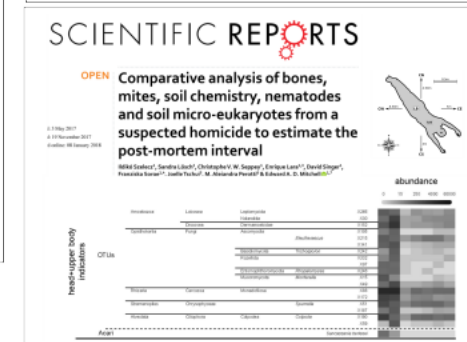


What's next for pr2-primers

- Update PR² to 5.1.0
- Include ROD database
- Add more primers
- ITS/28S primers

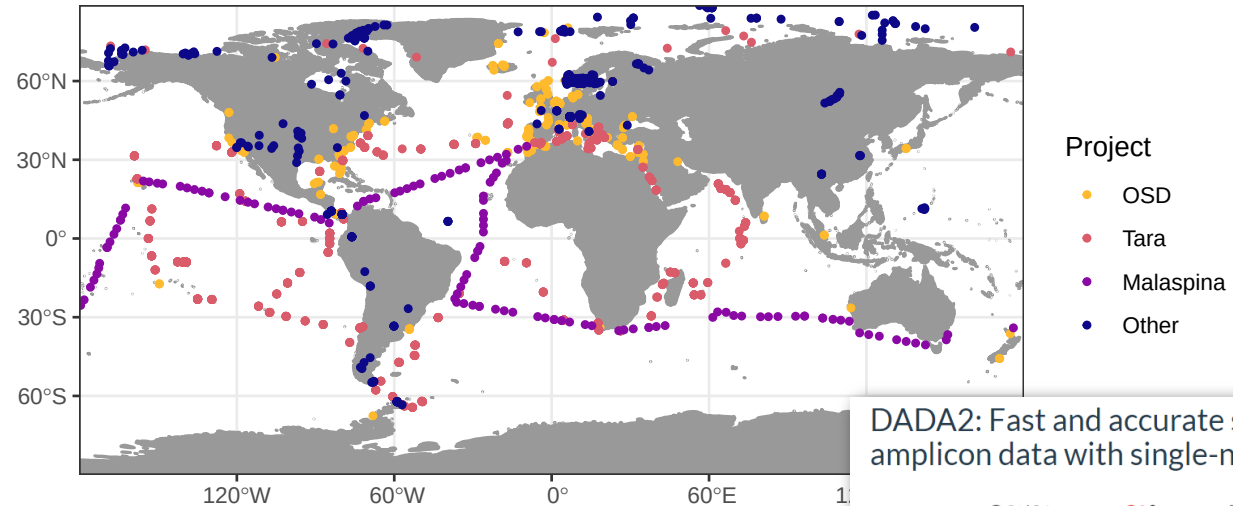


-
- Figure 1: Phylogenetic tree and relative abundance of microbial eukaryotes in geothermal springs. The tree is rooted at the bottom and branches upwards, showing relationships between various eukaryotic groups. A color scale on the right indicates relative abundance from 0.000 (white) to 0.001 (red). The tree is divided into five main clades: Metazoa (red), Opisthokonta (orange), Amoebozoa (yellow), Archaeplastida (green), and Alveolates (blue). The x-axis represents Temperature (°C) from 20 to 60. The y-axis represents Relative Abundance from 0.000 to 0.001. The tree shows that Alveolates and Archaeplastida are the most abundant groups, while Metazoa and Opisthokonta are less abundant. The tree also shows that some groups, like Amoebozoa, are more abundant at higher temperatures.



metapr2 - strategy

- Scan papers
- Start from raw data (fastq) available from GenBank SRA
- Use dada2 pipeline producing ASVs
 - Different datasets are comparable
- Annotate taxonomy with PR²
- Integrate metadata
 - Latitude and longitude
 - Depth
 - Substrate (water, ice, soil)
- Data stored in MySQL database
- Develop web interface using R shiny



Received: 7 February 2022 | Revised: 26 May 2022 | Accepted: 20 June 2022

DOI: 10.1111/1755-0998.13674

MOLECULAR ECOLOGY
RESOURCES WILEY

RESOURCE ARTICLE

metaPR²: A database of eukaryotic 18S rRNA metabarcodes with an emphasis on protists

Daniel Vault¹ | Clarence Wei Hung Sim² | Denise Ong² | Bryan Teo² |
Charlie Biwer³ | Mahwash Jamy³ | Adriana Lopes dos Santos²

DADA2: Fast and accurate sample inference from amplicon data with single-nucleotide resolution

DADA²
Amplicon Sequencing. Exactly. Version 1.18

metapr2 - version 2.1

- Datasets: 59

- ☐ Tara Oceans (reprocessed with dada2)
- ☐ Malaspina
- ☐ Ocean Sampling Day - 2014 & 2015
- ☐ Arctic datasets
- ☐ Deep Sea
- ☐ Lakes, Rivers, Soils

- Samples: 6,200

- ASVs: 90,000

- Annotated with PR2 version 5.0.1

<https://app.metapr2.org>

The metaPR2 database

Select datasets

41 items selected

dataset_id	dataset_name	region	paper_reference	sample_number	asv_number	n_reads_mean	selected
11	Antarctic Fields Bay-2013	Southern Ocean	Luo, W. et al. Molecular diversity of microbial eukaryotes in sea water from Fildes Peninsula, King George Island, Antarctica. Polar Biol. (2015)	10	69	13631	true
16	Antarctic Fields Bay 2015 18S V4	Southern Ocean	Trefaut, N., De la Iglesia, R., Moreno-Pino, M., Lopes dos Santos, A., G-e9-rikas Ribeiro, C., Parada-Pozo, G., Crist, A., Marie, D., & Vault, D. (2021) Annual phytoplankton dynamics in coastal waters from Fildes Bay, Western Antarctic Peninsula. Scientific Reports, 11(1), 1368.	123	685	48261	true
18	Antarctic Fields Bay 2015 18S V4 sorted	Southern Ocean	Trefaut, N., De la Iglesia, R., Moreno-Pino, M., Lopes dos Santos, A., G-e9-rikas Ribeiro, C., Parada-Pozo, G., Crist, A., Marie, D., & Vault, D. (2021) Annual phytoplankton dynamics in coastal waters from Fildes Bay, Western Antarctic Peninsula. Scientific Reports, 11(1), 1368.	60	280	31615	true

The metaPR2 database

Select datasets

41 items selected

Select Samples

Genes regions

V4

DNA or RNA

DNA

Ecosystems

oceanic, coastal, freshwater lakes, freshwater rivers, terrestrial

Substrates

water

Size fractions

pic, total

Depth levels

surface

Select ASVs

Minimum number of total reads per ASV

100

Select Taxa

Validate Taxa

Reset Taxa

Press VALIDATE after changing taxonomy to update screen.

Press RESET to reset taxonomy to top level (need to press validate after reset)

Supergroup

Archaeplastida

Division

Chlorophyta

Class

AS

Order

AS

Family

AS

Genus

AS

Species

Quick dataset selection.

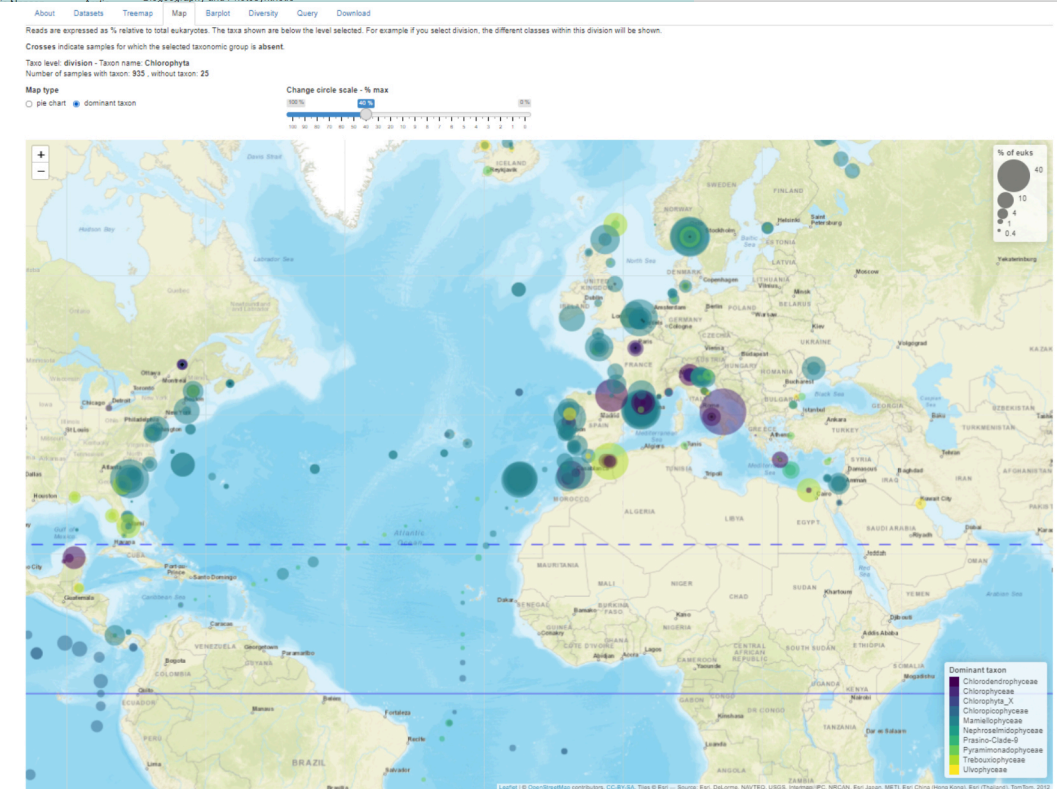
Dataset groups

☐ marine_global_V4 ☐ oceanic ☐ coastal ☐ rivers ☐ lakes ☐ soils ☐ arctic ☐ antarctic ☐ temperate ☐ tropical ☐ time series

Show 10 entries

Search:

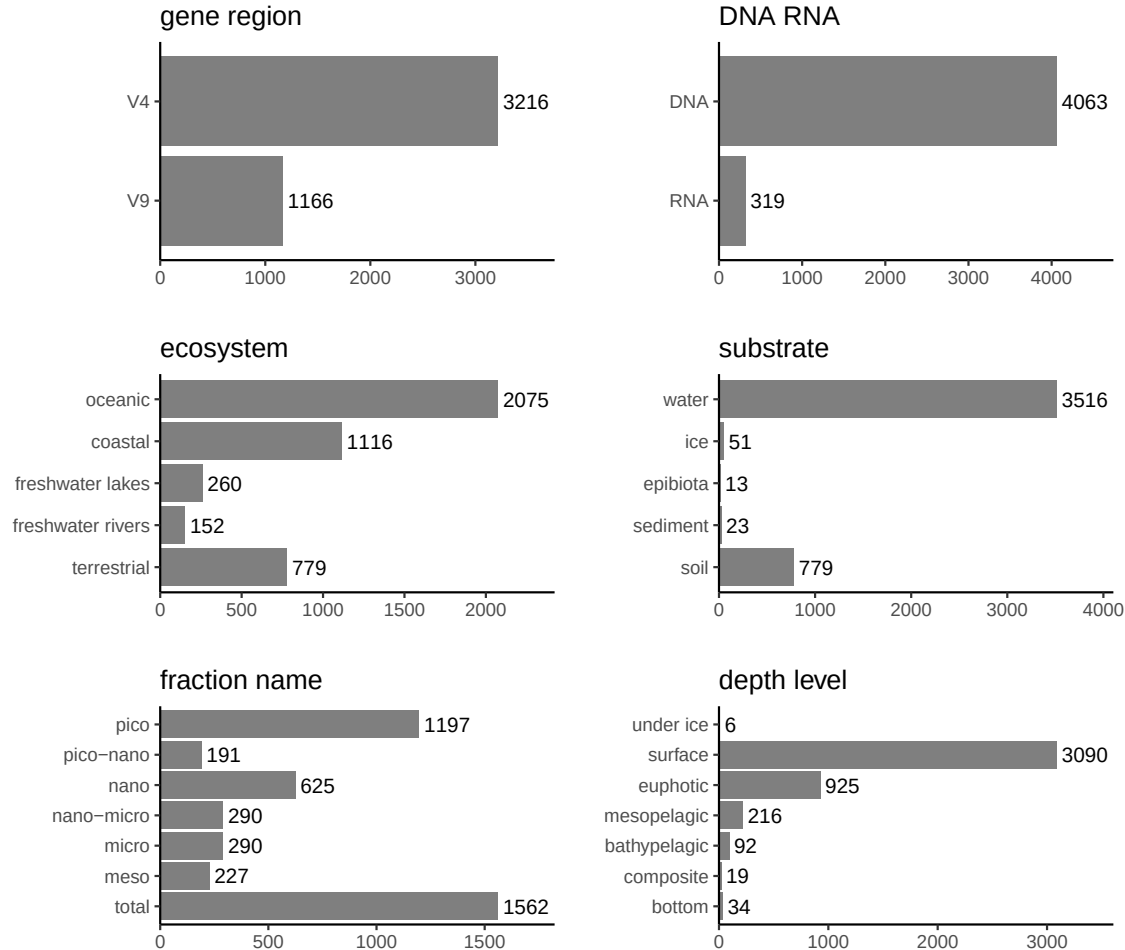
dataset_id	dataset_name	region	paper_reference	sample_number	asv_number	n_reads_mean	selected
11	Antarctic Fields Bay-2013	Southern Ocean	Luo, W. et al. Molecular diversity of microbial eukaryotes in sea water from Fildes Peninsula, King George Island, Antarctica. Polar Biol. (2015)	10	69	13631	true
16	Antarctic Fields Bay 2015 18S V4	Southern Ocean	Trefaut, N., De la Iglesia, R., Moreno-Pino, M., Lopes dos Santos, A., G-e9-rikas Ribeiro, C., Parada-Pozo, G., Crist, A., Marie, D., & Vault, D. (2021) Annual phytoplankton dynamics in coastal waters from Fildes Bay, Western Antarctic Peninsula. Scientific Reports, 11(1), 1368.	123	685	48261	true
18	Antarctic Fields Bay 2015 18S V4 sorted	Southern Ocean	Trefaut, N., De la Iglesia, R., Moreno-Pino, M., Lopes dos Santos, A., G-e9-rikas Ribeiro, C., Parada-Pozo, G., Crist, A., Marie, D., & Vault, D. (2021) Annual phytoplankton dynamics in coastal waters from Fildes Bay, Western Antarctic Peninsula. Scientific Reports, 11(1), 1368.	60	280	31615	true



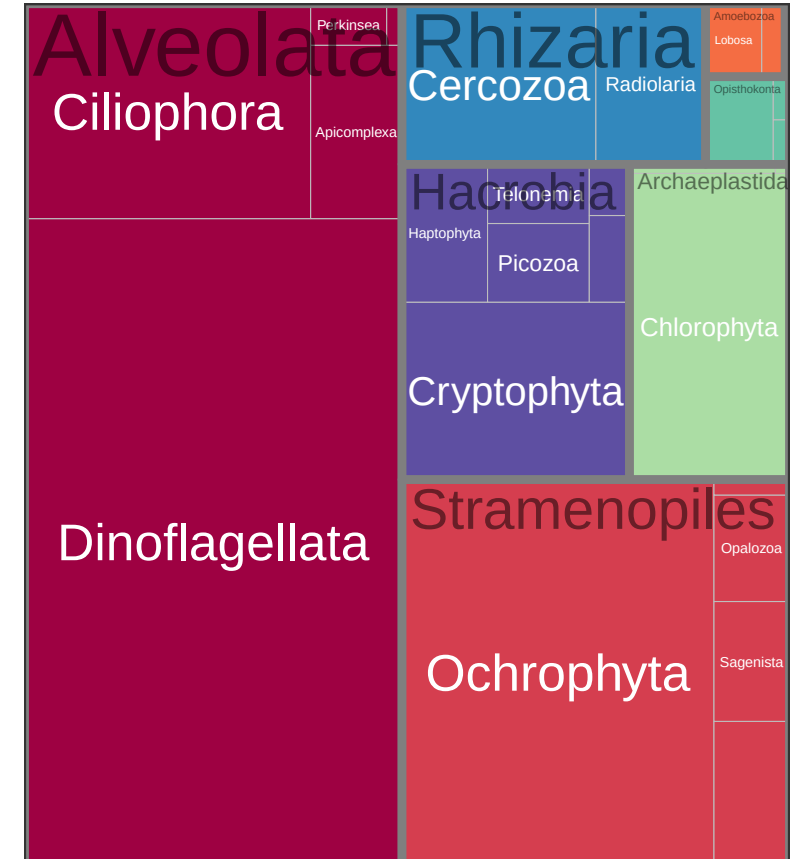
Samples and ASVs



Number of samples



Reads



Protists only

Web interface

<https://app.metapr2.org>



- Built with R shiny
 - Available also as R package
- Panels
 - Datasets
 - Treemaps
 - Maps
 - Barplots
 - Diversity
 - Query
 - Download


The metaPR2 database
[Help](#)

Select datasets
41 items selected

Select Samples
Gene regions
V4

DNA or RNA
DNA

Ecosystems
oceanic, coastal, freshwater lakes, freshwater rivers, terrestrial

Substrates
water

Size fractions
pico, total

Depth levels
surface

Select ASVs
Minimum number of total reads per ASV
100

Select Taxa
[Validate Taxa](#) [Reset Taxa](#)
Press VALIDATE after changing taxonomy to update screen.
Press RESET to reset taxonomy to top level (need to press validate after reset)

Supergroup
Archaeplastida

Division
Chlorophyta

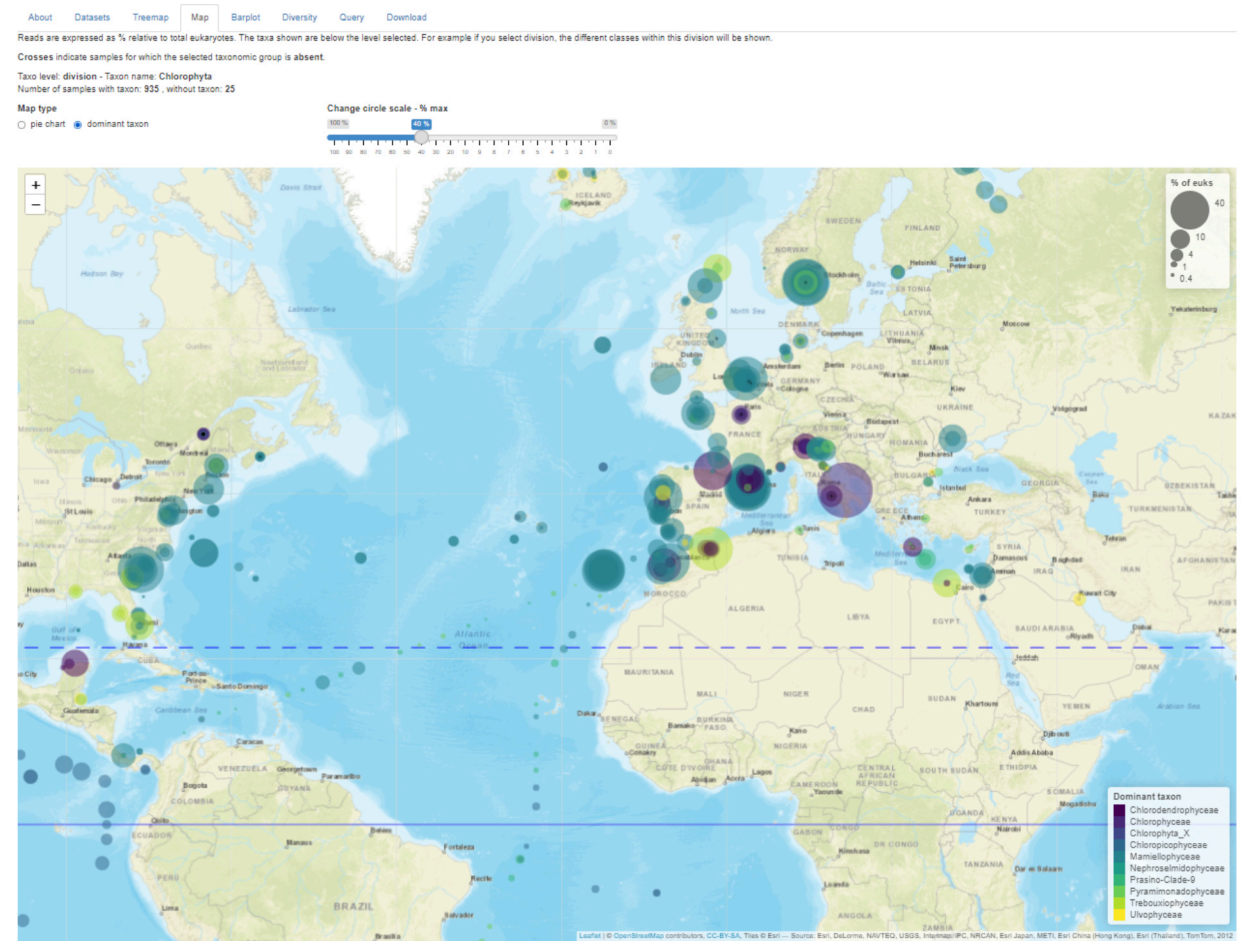
Class
All

Order
All

Family
All

Genus
All

Species

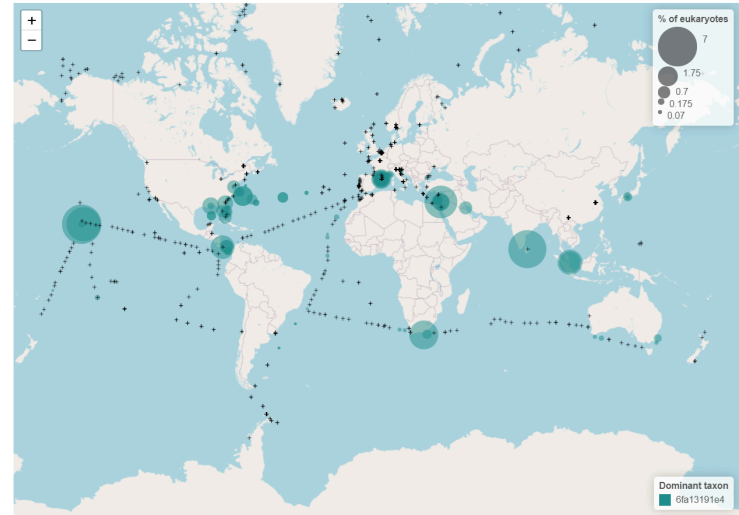
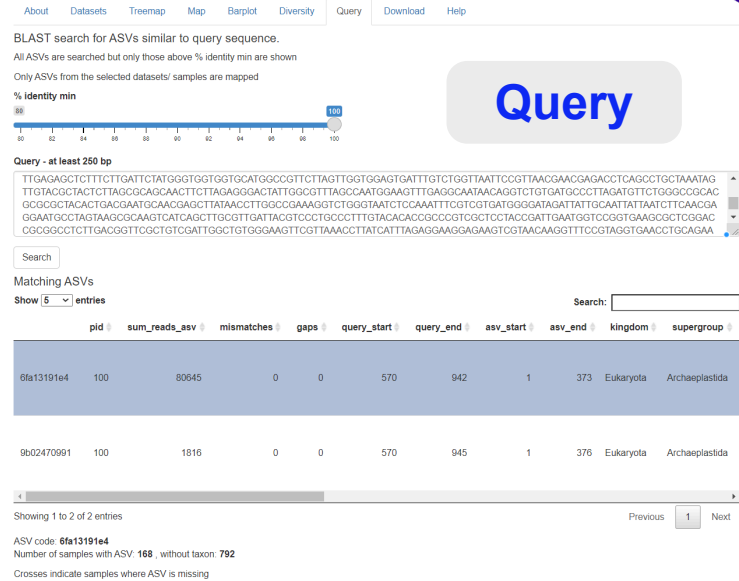




■ latitude ■ ecosystem ■ substrate ■ depth_level ■ depth ■ fraction_name ■ DNA_RNA ■ temperature ■ salinity
 Color
 ○ latitude ■ depth ○ temperature ○ salinity ■ fraction_name ○ substrate ○ ecosystem ○ depth_level ○ DNA_RNA

phyloseq-class experiment-level object
 site_label() 600 Factor [1200 taxa and 900 samples]
 sample_data() Sample Data: [900 samples by 9 sample variables]
 var_label() taxonomy Factor [1200 taxa by 9 taxonomic ranks]

Alpha Diversity Measures
 Chao1
 Shannon
 Simpson
 latitude



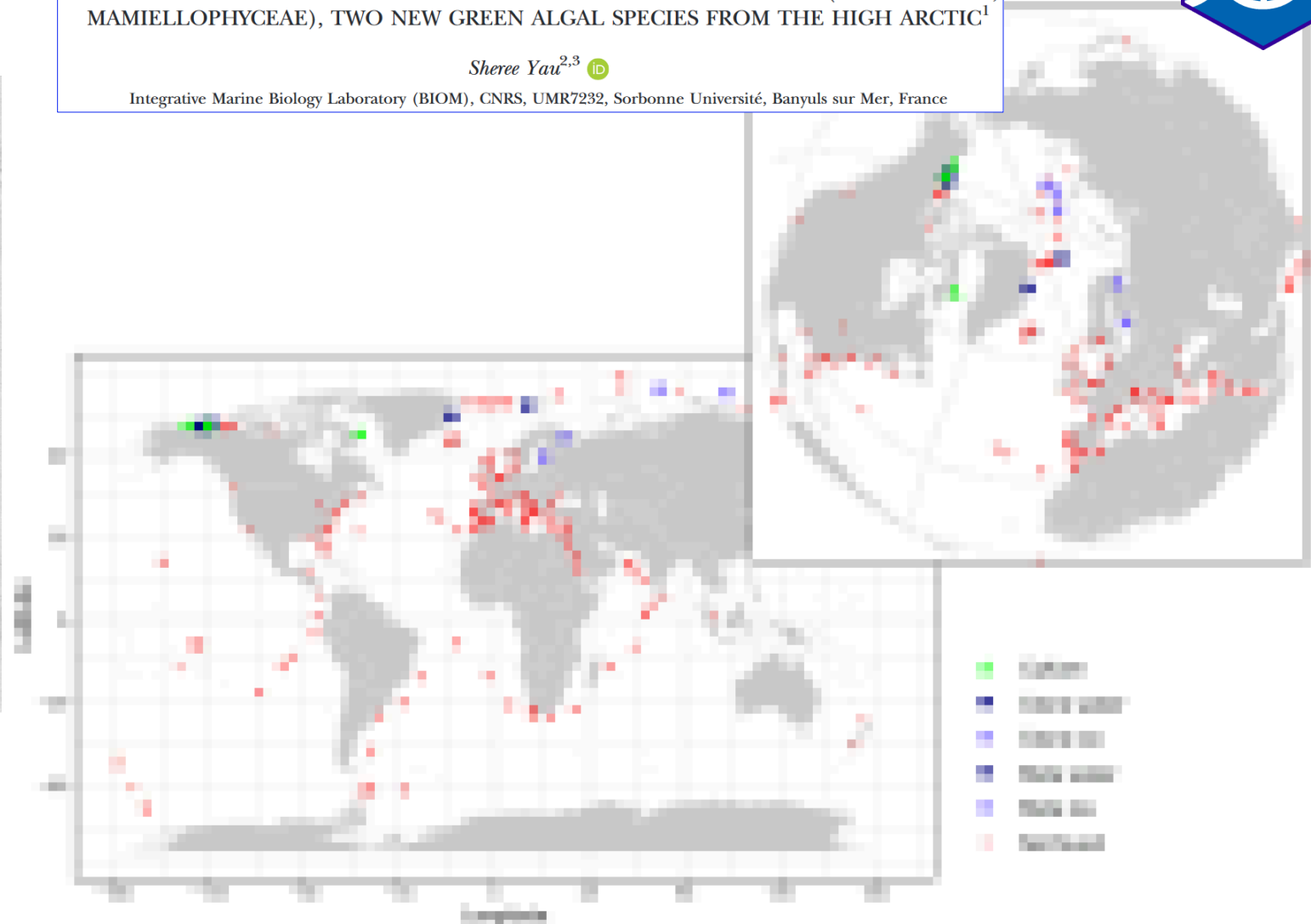
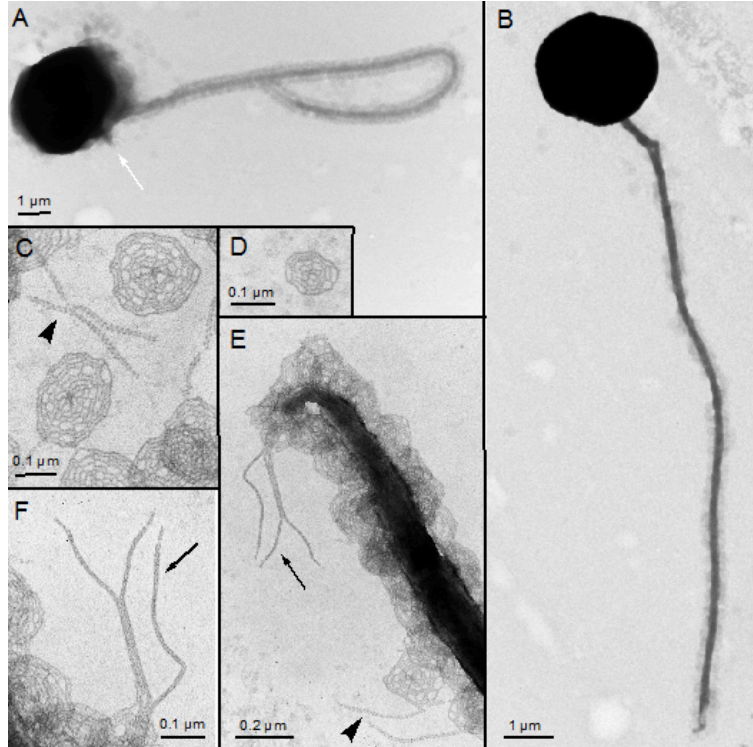
Biogeography

J. Phycol. **56**, 37–51 (2020)
© 2019 Phycological Society of America
DOI: 10.1111/jpy.12932

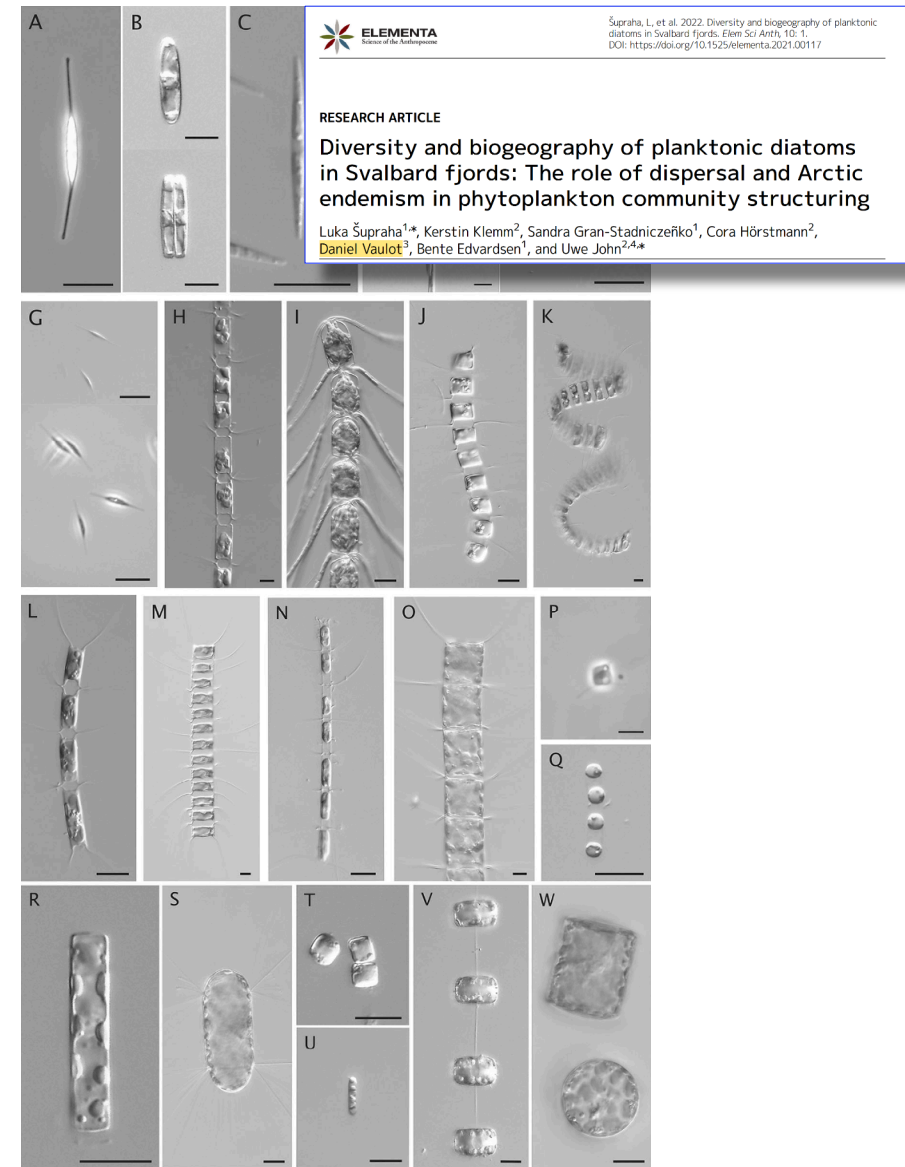
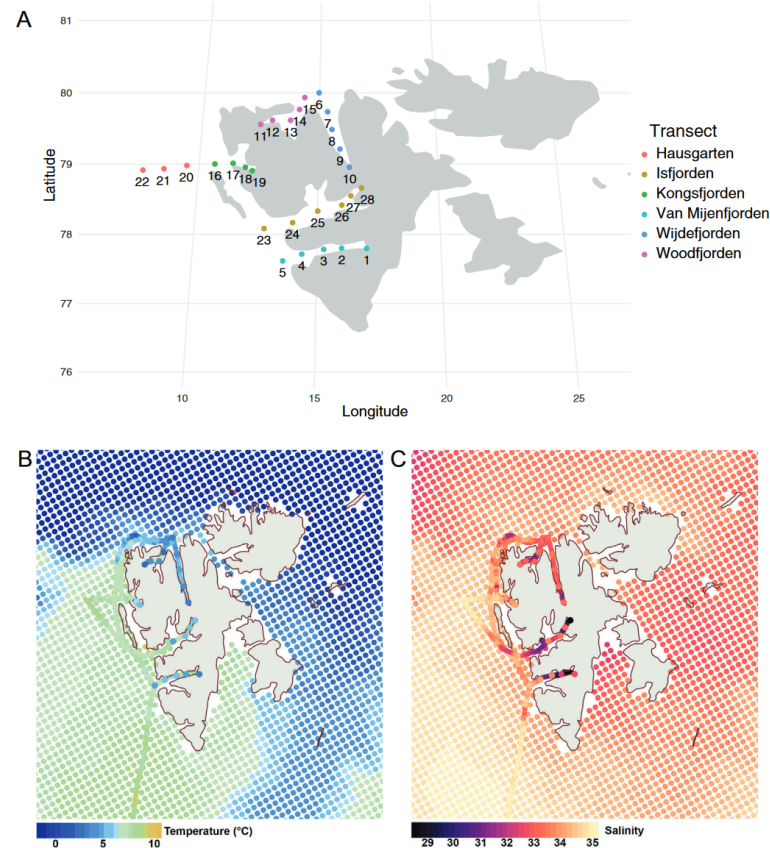
MANTONIELLA BEAUFORTII AND *MANTONIELLA BAFFINENSIS* SP. NOV. (MAMIELLALES, MAMIELLOPHYCEAE), TWO NEW GREEN ALGAL SPECIES FROM THE HIGH ARCTIC¹

Sheree Yau^{2,3} 

Integrative Marine Biology Laboratory (BIOM), CNRS, UMR7232, Sorbonne Université, Banyuls sur Mer, France



Svalbard diatoms



Šupraha, L. et al., 2022. Diversity and biogeography of planktonic diatoms in Svalbard fjords: the role of dispersal and Arctic endemism in phytoplankton community structuring. *Elementa: Science of the Anthropocene* 10, 00117

Biogeographic distribution types

RESEARCH ARTICLE

Diversity and biogeography of planktonic diatoms in Svalbard fjords: The role of dispersal and Arctic endemism in phytoplankton community structuring

Luka Šupraha^{1,*}, Kerstin Klemm², Sandra Gran-Stadniczeŕko¹, Cora Hörstmann²,
Daniel Vaulot³, Bente Edvardsen¹, and Uwe John^{2,4,*}

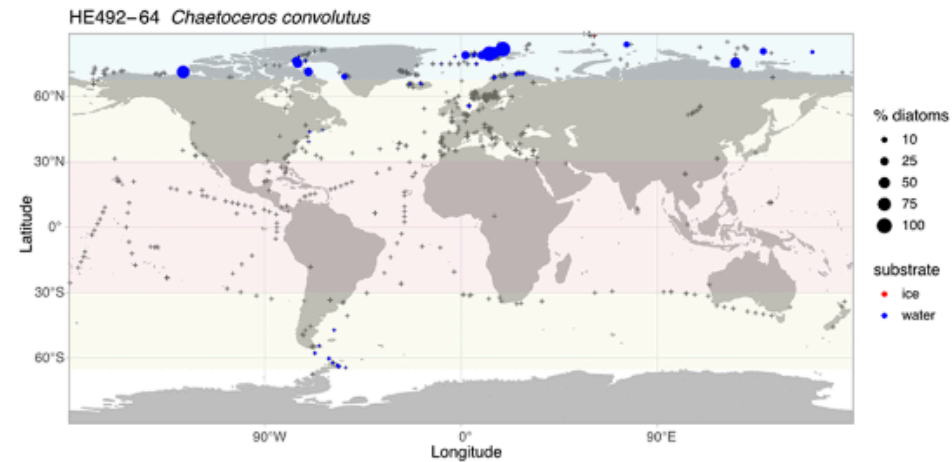
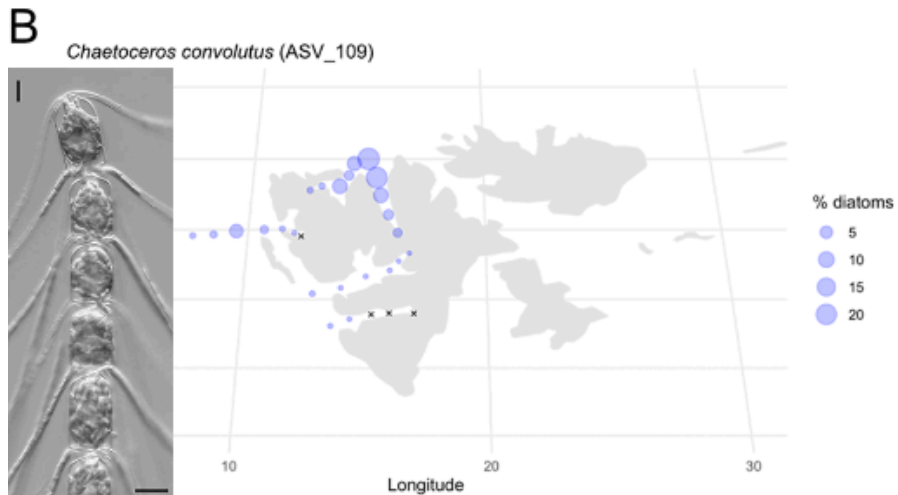
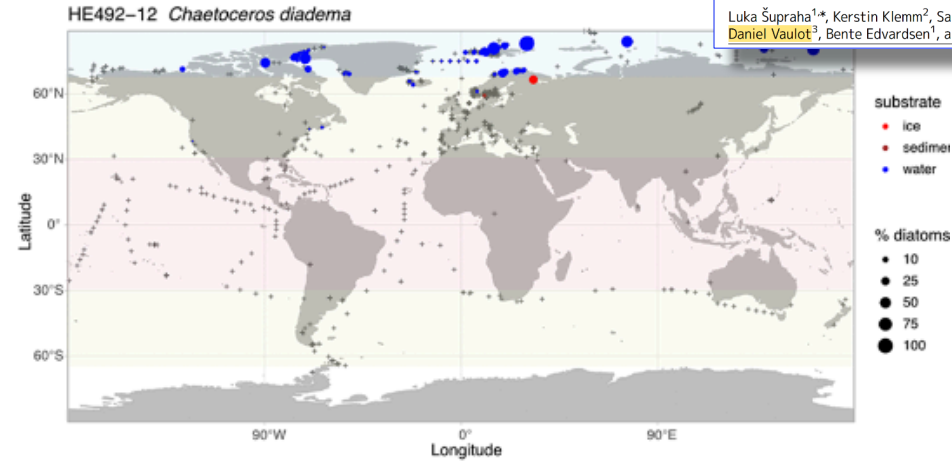
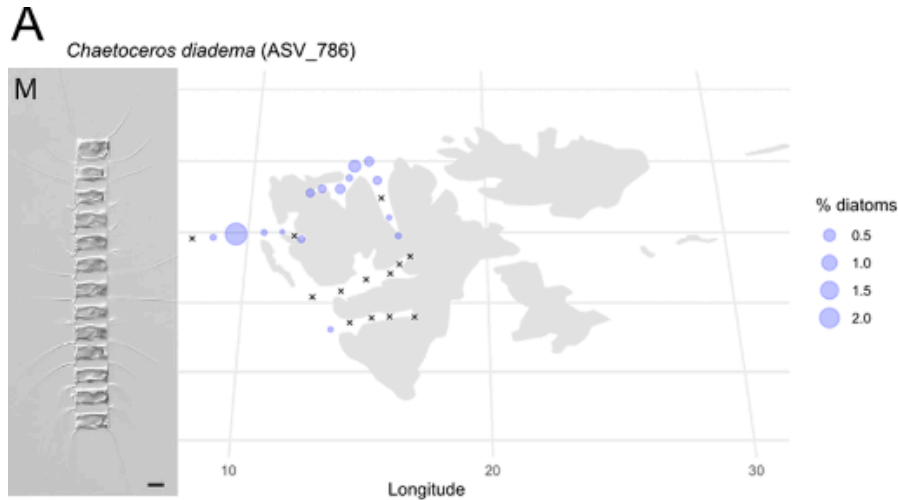
Biogeographic Distribution Type	Description
Arctic	Genotype has a biogeographic distribution mostly restricted to the Arctic circle above 66°N
Arctic-temperate	Genotype is present in the Arctic and at temperate latitudes 30°N–66°N and 30°S–66°S
Cosmopolitan	Genotype is present in the Arctic and has a cosmopolitan distribution

Polar genotypes

RESEARCH ARTICLE

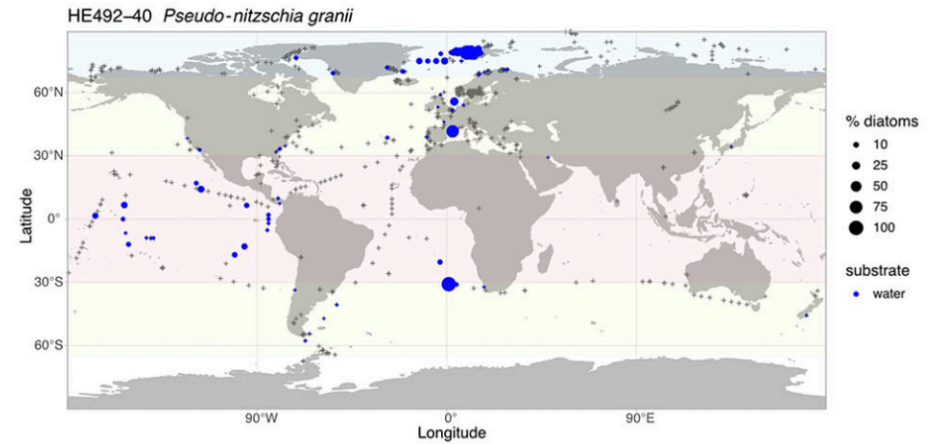
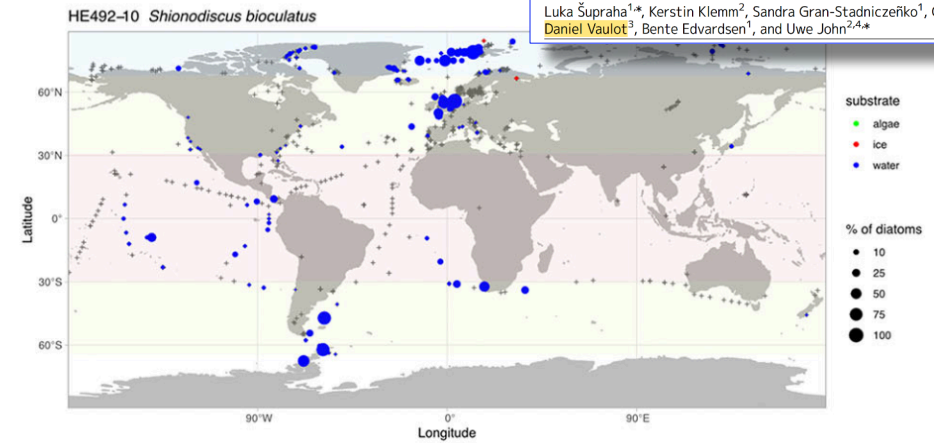
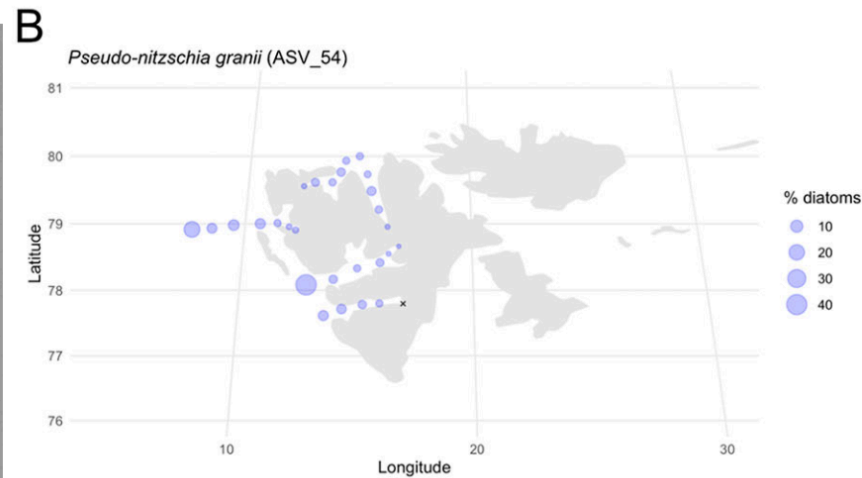
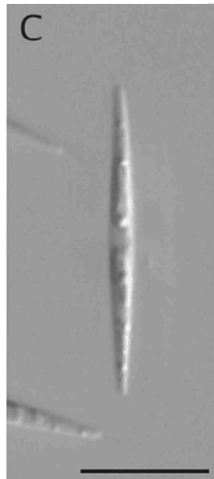
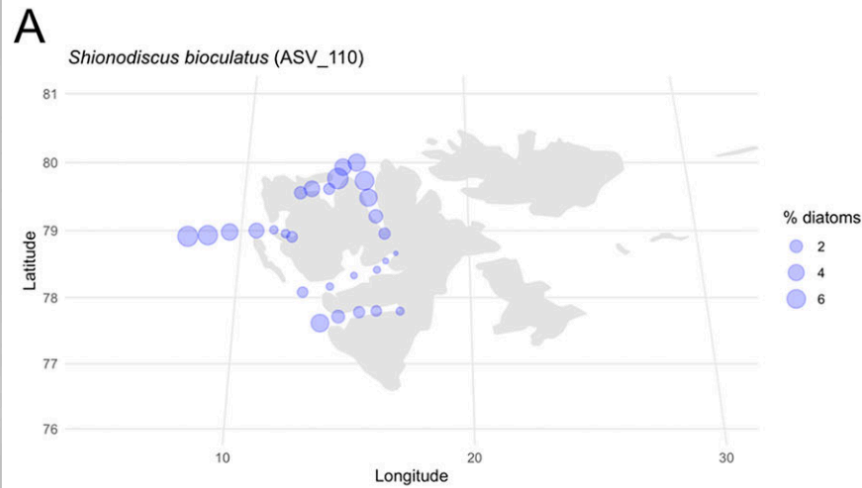
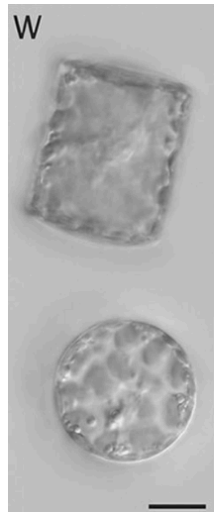
Diversity and biogeography of planktonic diatoms in Svalbard fjords: The role of dispersal and Arctic endemism in phytoplankton community structuring

Luka Šupraha^{1,*}, Kerstin Klemm², Sandra Gran-Stadniczeŋko¹, Cora Hörstmann²,
Daniel Vaulot³, Bente Edvardsen¹, and Uwe John^{2,4,*}



Šupraha, L. et al., 2022. Diversity and biogeography of planktonic diatoms in Svalbard fjords: the role of dispersal and Arctic endemism in phytoplankton community structuring. Elementa: Science of the Anthropocene 10, 00117

Cosmopolitan genotypes

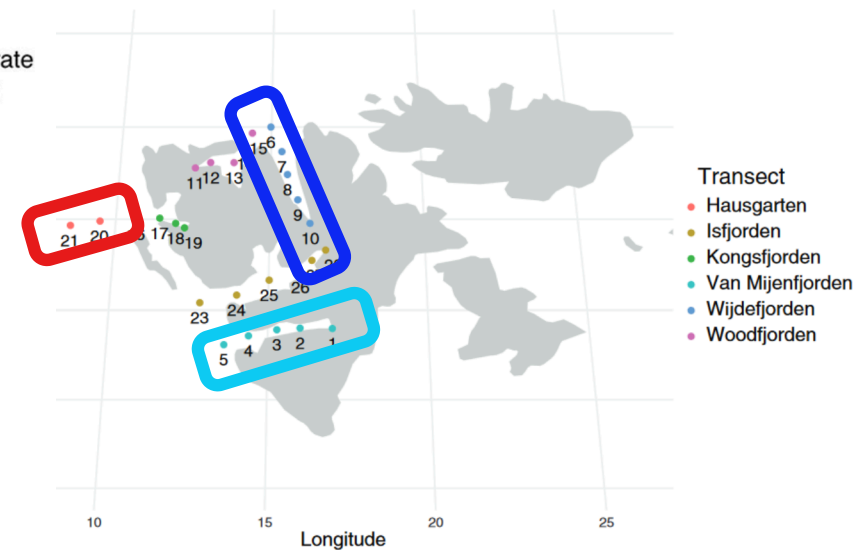
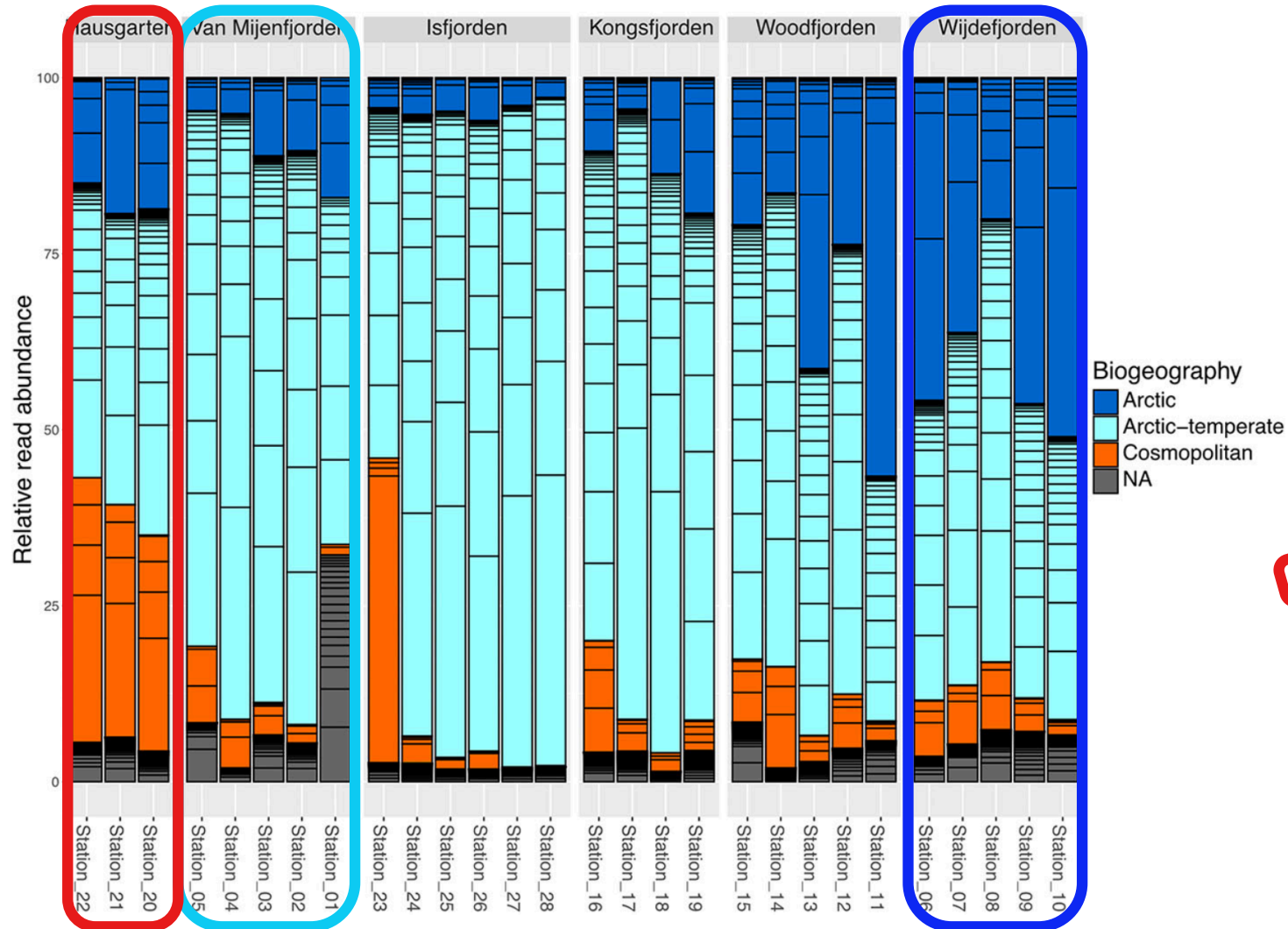


RESEARCH ARTICLE

Diversity and biogeography of planktonic diatoms in Svalbard fjords: The role of dispersal and Arctic endemism in phytoplankton community structuring

Luka Šupraha^{1,*}, Kerstin Klemm², Sandra Gran-StadniczeŹko¹, Cora Hörstmann²,
Daniel Vaulot³, Bente Edvardsen¹, and Uwe John^{2,4,*}

Distribution in Svalbard fjords



RESEARCH ARTICLE

Diversity and biogeography of planktonic diatoms in Svalbard fjords: The role of dispersal and Arctic endemism in phytoplankton community structuring

Luka Šupraha^{1,*}, Kerstin Klemm², Sandra Gran-StadniczeŹko¹, Cora Hörstmann², Daniel Vaulot³, Bente Edvardsen¹, and Uwe John^{2,4,*}

What's next for **metapr2**

- More datasets
 - antarctic, tropical, equatorial
 - soils, sediments
 - freshwater
 - microbiomes
- Reannotate with PR² version 5.1
- Include long read metabarcodes ?

Thanks to all PR² contributors



Contributors PR2 reference



Javier del Campo
Institute of Evolutionary Biology
UPF-CSIC
PR2 reference, EukRef



Mahwash Jamy
Swedish University of
Agricultural Sciences, Uppsala
Eukaryotic phylogeny



Anna Maria Fiore-Donno
Universität zu Köln
Amoebozoa



Raphaël Morard
Marum, University of Bremen
Foraminifera



Miguel Mendez Sandin
Uppsala U., Sweden
Radiolaria



Alexandra Schönlé
Universität zu Köln
Stramenopiles



Laure Arsenieff
CNRS, France
Diatoms



Ana Maria Cabello
IEO-Málaga
Pelagophyceae



Charles Bachy
CNRS, France
Ciliates



Tristan Biard
U. Littoral, France
Radiolaria



Vittorio Boscaro
UBC, Canada
Ciliates



Fabien Burki
Uppsala U., Sweden
Eukaryotic phylogeny



Andrea de Oliveira da Rocha Franco
FURG, Brazil
Diatoms



Johan Decelle
CEA, France
Plastids



Richard Dorrell
Institut de biologie de l'École
normale supérieure, Paris
Plastid 16S



Manon Dünn
Universität zu Köln, Germany
Percolomonads



Bente Edvardsen
U. Oslo, Norway
Haptophyta



Maximilian Ganser
Paris-Lodron University of
Salzburg, Austria
Ciliates



Chetan Gaonkar
Texas A&M, USA
Diatoms



Megan Gross
University of Kaiserslautern,
Germany
Microsporidia



Contributors PR2 primers



Frédéric Mahé
CIRAD, France
Bioinformatics



David Bass
CEFAS, UK
Parasitic protists



Stefan Geisen
Wageningen U., The Netherlands
Soil protists



Daniel Vaultot
CNRS, France
PR2 coordinator, metaPR2



Javier del Campo
Institute of Evolutionary Biology
UPF-CSIC
PR2 reference, EukRef

Scientific Committee



Frédéric Mahé
CIRAD, France
Bioinformatics



Fabien Burki
Uppsala U., Sweden
Tree of eukaryotes



Bente Edvardsen
U. Oslo, Norway
Haptophyta



Stefan Geisen
Wageningen U., The Netherlands
Soil protists



Laure Guillou
CNRS, France
PR2 reference



Martin Kolisko
CAS, Czech Republic
Apicomplexa



Adriana Lopes dos Santos
NTU, Singapore
Marine protists



Ramon Massana
ICM, Spain
Stramenopiles



Fabrice Not
CNRS, France
Radiolaria



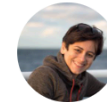
Laura Parfrey
UBC, Canada
Protists



Ian Probert
Sorbonne Université, France
Roscoff Culture Collection



Luciana Santoferrara
Hofstra University, USA
Ciliates



Adriana Lopes dos Santos
Visiting Professor
Assistant Professor
📧 🌐 📱 📺 📺



Daniel Vaultot
Assistant Professor
📧 🌐 📱 📺 📺



Fabien Burki
Uppsala U., Sweden

PhD students



Clarence Sim
PhD student



Denise Ong
PhD student



Bryan Teo
URECA Undergraduate student



Mahwash Jamy
Uppsala U., Sweden



Haifeng Gu
Third Institute of Oceanography,
China
Dinoflagellates



Adriana Lopes dos Santos
NTU, Singapore
Marine protists



Ramon Massana
ICM, Spain
Stramenopiles



Kenneth Mertens
IFREMER, France
Dinoflagellates



Sebastian Metz
Station Biologique, Roscoff
Perkinsids



Solenn Mordret
SZN, Italy
Dinoflagellates



Luciana Santoferrara
Hofstra University, USA
Ciliates



Se Hyeon Jang
Chonnam National University,
Korea
Dinoflagellates



Алексей Селюк
Saint Petersburg State University
Ciliates



Pavel Škaloud
Charles University, Prague, CZ
Chrysophyceae



Margot Tragin
IFREMER, France
Chlorophyta



Wei-Ting Chen
NTO U., Taiwan
Ciliates

Web site: <https://pr2-database.org/>

Moving from Slack: <https://matrix.to/#/#pr2:matrix.org>

The PR² databases

About Download Documentation Annotation Workshops Papers News

The PR² databases

Three interconnected 18S rRNA databases

- PR2 reference sequence database
- PR2 primers
- metaPR2 metabarcodes

The PR² database ecosystem

The eukaryotic tree. From Burki et al. 2019. The New Tree of Eukaryotes. Trends in Ecology & Evolution. DOI: 10.1016/j.tree.2019.08.008.

The PR² (Protist Ribosomal Reference) database ecosystem is a set of three interconnected 18S rRNA databases that are useful in particular for metabarcoding applications. This web site focuses on the PR² reference sequence database.

The PR² reference sequence database

- Current version : 4.14.0
- Last update : 25 June 2021
- DOI [10.5281/zenodo.5031733](https://doi.org/10.5281/zenodo.5031733)
- Download: <https://github.com/pr2database/pr2database/releases>
- [downloads 228](#)
- Contributors

Element | PR2 announcements and questions

Search Ctrl K

pr2

Favourites

PR2 announcements and questions

People

Javier del Campo

luciana.santoferrara

Adriana lopes dos santos

Fabien

Rooms

PR2-primers

History Dear Dr. Vault, I'm a PhD-student from ...

PR2 taxo Stramenopiles

Thank you Andrea de Oliveira da Rocha Franco .

PR2 Functional traits

Files PR2 traits 2022-05-23.pdf; presentation of ...

PR2 taxo Fungi and related

Aneceai Canoc. Okay, got it

PR2 taxo Alveolates other

Sebastian Metz: Hi Sebastian, here are the Perk...

PR2 taxo 16S plastid

History Hieroen Verbruggen 6 h 23 @Daniel Vaul...

PR2 Full open

Javier del Campo: Great!

PR2 taxo dinoflagellates

halfeungu: Dear colleagues, can you add these ...

PR2 taxo Amoebozoa

Files for Myxomycetes annotations

PR2 taxo Ciliates

For those of you that volunteered for annotation...

Show less

PR2 announcements and questions

The Protist Ribosomal Reference database of eukaryotic rRNA sequences. <https://pr2-database.org/> Please ask here any question about PR2, pr2-primers or metapr2

The main changes are as follows

- New database: version 2.0 - 59 datasets (18 new)
- Clustering: An option is now provided to use either all ASVs or clustered ASVs on the welcome screen. ASVs are clustered at 100% identity with VSEARCH --id 1.00 See the metaPR2 paper (<https://doi.org/10.1111/1755-0998.13674>) for more information.
- It is also possible to use version 1.0 of the database by entering 'v1' on the welcome screen
- Taxonomy: This new panel provides a table with all the taxa present in the current metaPR2 version with the number of ASV for each species. The table can be easily searched.
- Taxonomy is constructed from all the samples and not only samples selected.
- Barplots: The right side of the graph indicates, for each parameter range, the number of samples that fall into that range as well as the number of samples that contain the taxa selected.

In addition, if you have some strong request for new features let me know.

A big thanks in advance. Daniel
(edited)

MetaPR2

English Choose datasets Datasets ('blank' for version 2.0 and 'v1' for version 1.0): Password ('blank' for version 1.0 and 2.0):

Monday

pavel.skalous

Daniel Vaultot

@room

Dear all...

Dear Daniel, thank you for such an amazing effort! I am still new to PR2 - I tried to check the Ochrophyte-Chrysophyte databases and it seems there is still an older taxonomy (Synurophyceae, Chrysophyceae_X as a single order, and so on). Yours, Pavel

Daniel Vaultot

pavel.skalous: Dear Pavel.

Thanks for checking metaPR2. The taxonomy used for metaPR2 is still that of PR2 4.14.0. I am planning an update of PR2 (4.15.0) hopefully before the end of the year and this will include the Chrysophyceae update that you provided. Once PR2 is updated then I will update metaPR2... This is long process with small steps.

Tuesday

pavel.skalous

Daniel Vaultot

@room

Dear Pavel.

Thanks for checking metaPR2. The taxonomy used for metaPR2 is still that of PR2 4.14.0. I am planning an update of PR2 (4.15.0) hopefully before the end of the year and...

Great to know, thanks :).

Daniel Vaultot

@room

An paper relevant to PR2 management just came out: <https://onlinelibrary.wiley.com/doi/abs/10.1111/1755-0998.13723>

Management of DNA reference libraries for barcoding and metabarcoding studies with the R package refdb - Wiley Online Library

DNA barcoding and metabarcoding are revolutionizing the study and survey of biodiversity. In order to assign taxonomic labels to the DNA sequence data retrieved, these methods are strongly dependent ...

Send a message...